

Using Genomic Databases for Sequence-Based Biological Discovery

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The inherent potential underlying the sequence data produced by the International Human Genome Sequencing Consortium and other systematic sequencing projects is, obviously, tremendous. As such, it becomes increasingly important that all biologists have the ability to navigate through and cull important information from key publicly available databases. The continued rapid rise in available sequence information, particularly as model organism data is generated at breakneck speed, also underscores the necessity for all biologists to learn how to effectively make their way through the expanding "sequence information space." This review discusses some of the more commonly used tools for sequence discovery; tools have been developed for the effective and efficient mining of sequence information. These include LocusLink, which provides a gene-centric view of sequence-based information, as well as the 3 major genome browsers: the National Center for Biotechnology Information Map Viewer, the University of California Santa Cruz Genome Browser, and the European Bioinformatics Institute's Ensembl system. An overview of the types of information available through each of these front-ends is given, as well as information on tutorials and other documentation intended to increase the reader's familiarity with these tools.

INTRODUCTION

In April 2003, the scientific community celebrated the achievement of the Human Genome Project's major goal: completion of a high-accuracy sequence of the human genome. The significance of attaining this goal, which many have compared with landing a man on the moon, cannot be underestimated. This milestone firmly marks the entrance of modern biology into the genomic era (and *not* the post-genomic era, as many have stated), changing the way in which biological and clinical research will be conducted in the future. The intelligent use of sequence data from human and model organisms, along with technological innovations fostered by the Human Genome Project, will lead to significant advances in our understanding of diseases that have a genetic basis and, more importantly, in how health care is delivered from this point forward.

The completion of human genome sequencing has provided the biological community an opportunity to look forward and begin to think about how to use genomic approaches in a way that will lead to tangible health benefits. To that end, the National Human Genome Research Institute led a 2-year process involving hundreds of scientists and members of the public in more than a dozen workshops and individual consultations. The result of this process has led to the publication of a document entitled *A Vision for the Future of Genomics Research* (1). This "vision document" sets forth a number of "grand challenges" organized around 3 major themes: genomics to biology, genomics to health, and genomics to society. These grand challenges are intended to provide ambitious, interdisciplinary research goals for the scientific community that will eventually translate the promise of the Human Genome Project into improved human health.

As part of this vision, 6 critical "cross-cutting elements" were identified as being relevant to all 3 of the thematic areas. One of these areas is computational biology, an area whose importance will continue to increase as more and more sequence data becomes available, as data sets continue to get larger and larger, and as the complexity of both the data and the kinds of questions being addressed become more sophisticated. The focus on computational biology (or, as it is more often called, bioinformatics) underscores that both laboratory- and computationally-based approaches will be necessary to do cutting-edge research in the future. In the same way that investigators are trained in basic biochemistry and molecular biology techniques, a basic understanding of bioinformatic techniques as part of the biologist's arsenal will be absolutely indispensable in the future.

The database that most biologists are familiar with is GenBank (2), an annotated collection of all publicly available DNA and protein sequences maintained by the National Center for Biotechnology Information (NCBI) at the National Institutes of Health. At the time of this writing, GenBank contained 35.6 billion nucleotide bases, representing 29.8 million sequences in more than 119000 species (2). Whereas the inherent value of these data cannot be understated, the sheer magnitude of data presents a conundrum to the inexperienced user, not just because of the size of the "sequence information space," but because the information space continues to get larger, growing at an exponential pace. GenBank's size doubles once every 12 to 14 mo; this translates to 45 new sequences being deposited every minute and 7 new structures becoming available every day. This exponential growth rate is expected to continue well into the future, particularly because of the September 2002 announcement of "high priority" model organisms earmarked for

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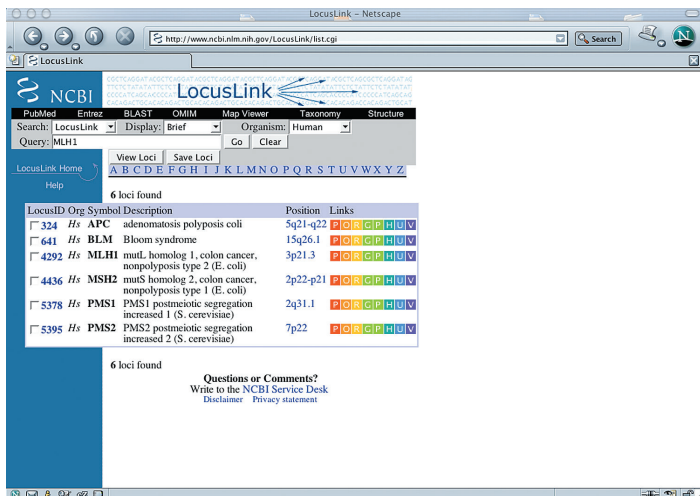


Figure 1. Results of a LocusLink query, using “MLH1” as the search term. The report returns information on the MLH1 gene, as well as for related genes. A brief description is given for each found locus, as well as its chromosomal location. The colored alphabet blocks found to the right of each entry are explained in detail in the text.

sequencing. The continued, rapid rise in available sequence information underscores the necessity for all biologists to learn how to effectively make their way through this sequence space. GenBank (or any other biological database, for that matter) serves little purpose unless the data can be easily searched and entries retrieved in a usable, meaningful format. Otherwise, sequencing efforts will have had no useful end, because the biological community as a whole would not be able to make use of the information hidden within these millions of bases and amino acids. Much effort has gone into making these data available to the biological community, and several of the most highly used interfaces resulting from these efforts are the focus of this review.

GENE-CENTRIC INFORMATION RETRIEVAL

One of the most commonly used interfaces for retrieving biological data is LocusLink, which allows access to a series of component databases in a gene-centric fashion (3). The kinds of information that can be retrieved using LocusLink include nucleotide and protein sequences, bibliographic information from PubMed, information on homologous genes, and single nucleotide polymorphism (SNP) data. LocusLink queries are very easy to perform, requiring only the name of the gene of interest or the gene symbol. As an example, we will search for information on the gene MLH1. From the LocusLink home page (<http://www.ncbi.nlm.nih.gov/LocusLink/>), the user would simply type “MLH1” into the query box, select “Human” as the organism, then click *Go*. The results of the query are shown in Figure 1. Note that MLH1 is the 3rd entry in the returned list; the other entries returned are also related to MLH1 and may be of interest to the user. Clicking on the LocusID number at the left of the entry (here, 4292) returns some basic information about the gene, giving the official gene symbol and name, as well as a brief summary of the function of the gene.

The colored alphabet blocks that appear to the right of each entry are hyperlinked and allow the user to find all available

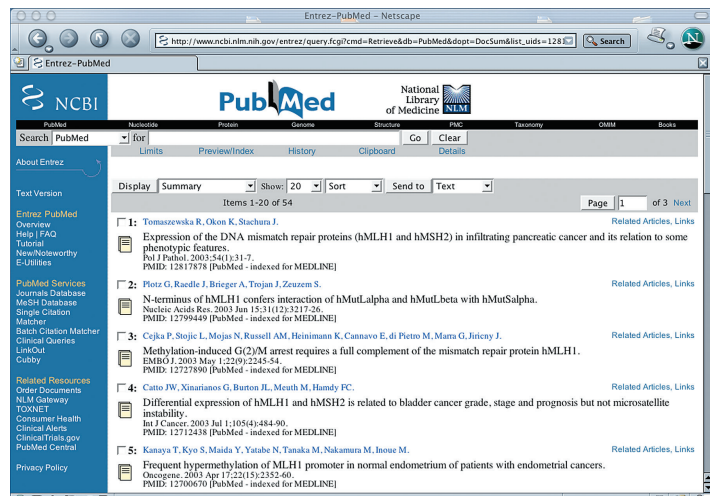


Figure 2. A PubMed window showing papers on MLH1. Each entry gives the names of the authors, the title of the paper, and the citation information. The abstract of each paper can be found by clicking on the hyperlinked list of authors.

information on the particular gene of interest. Clicking on the 1st *P* (in red) in the row marked MLH1 takes the user to PubMed, the National Library of Medicine’s collection of citations from the primary literature (Figure 2). The list is arranged in reverse chronological order (most recent 1st) and clicking on the hyperlinked author lists returns the abstract for that particular paper. Also notice the *Related Articles, Links* hyperlink to the right of each author list; clicking on these links returns a list of all of the papers that are similar in subject matter to the original paper.

Returning to the original hit list, clicking the *O* block takes the user to Online Mendelian Inheritance in Man (OMIM), an often-overlooked but incredibly valuable resource for anyone studying genetic or genomic disorders (4). OMIM provides concise textual information from the published literature on most human conditions having a genetic basis, as well as pictures illustrating the condition or disorder (where appropriate) and full citation information (Figure 3A). One of the most important features of OMIM is that for each gene, a list of allelic variants is available (see Figure 3B). Clicking on any of the items in the list returns information on how specific mutations (shown in the square brackets at the end of each entry) were identified and the effect of those specific mutations in patients. In this case, a number of allelic variants have been identified in the MLH1 gene leading to hereditary nonpolyposis colon cancer, but also related to some other syndromes as well.

The next 3 alphabet blocks would take the user to actual sequence information for that gene. The *R* stands for RefSeq, and clicking on the *R* would take the user to the “reference sequence” for that entry. The RefSeq project at NCBI is geared toward reducing redundancy in the public databases, with the goal of representing each molecule in the central dogma (DNA, mRNA, or protein) by 1 and only 1 sequence. Often times, a user will do a query and get back a long list of sequences, all representing the same biological entity, and it is often unclear which entry should be used; by using the curated RefSeq entry, the user can be assured that they are using the most accurate sequence information available. The *G*

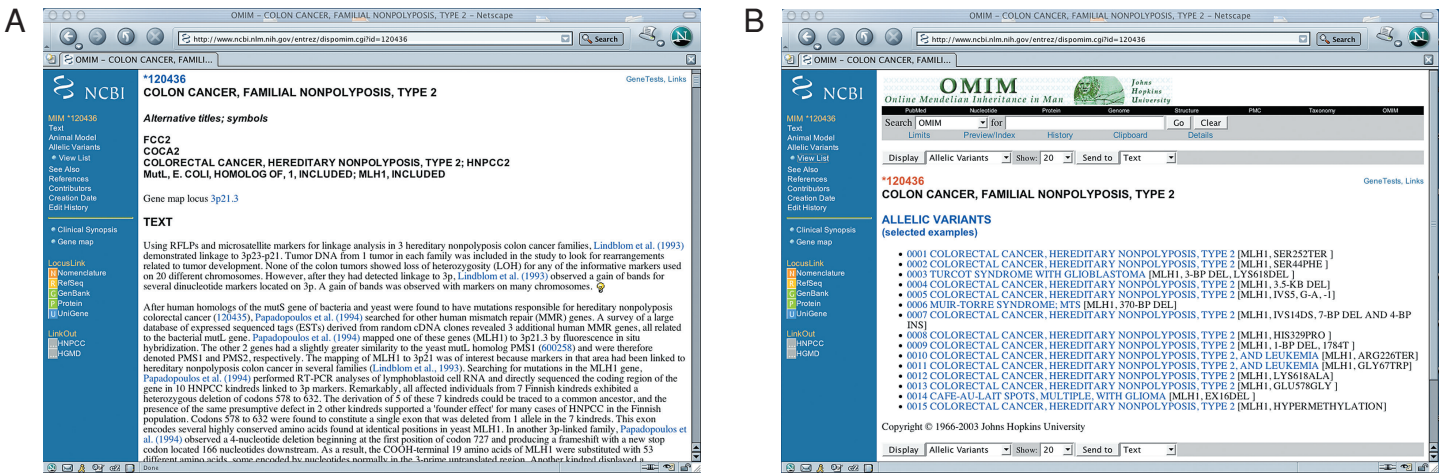


Figure 3. Information from Online Mendelian Inheritance in Man (OMIM) for the MLH1 gene. (A) OMIM entries begin with general information on the gene; the section marked *Text* provides an “executive summary” of relevant information on the gene and is curated on a regular basis by experts in the field. (B) For each gene where information is available, a list of allelic variants can be obtained; clicking on any of the entries provides more detailed information on that particular variant. See text for details.

and *P* are for GenBank and Protein, respectively, and will return *all* nucleotide and protein entries available for the gene of interest.

The remaining 3 alphabet blocks take users to more specialized resources, and for purposes of this discussion, we will take them slightly out of order, beginning with the *U*. The *U* stands for UniGene, a long-standing effort at NCBI to collapse expressed sequence tags (ESTs) (5), mRNA sequences, and coding sequences into discrete clusters. All of the sequences represented in a particular UniGene cluster share similarity at their 5' end, and the presence of at least 1 EST in each cluster means that, by definition, the cluster represents the transcription product of a distinct gene. An example of a UniGene entry page, found by clicking on the *U* resulting from the original query, can be found in Figure 4. The page begins with information on selected model organism protein similarities, showing

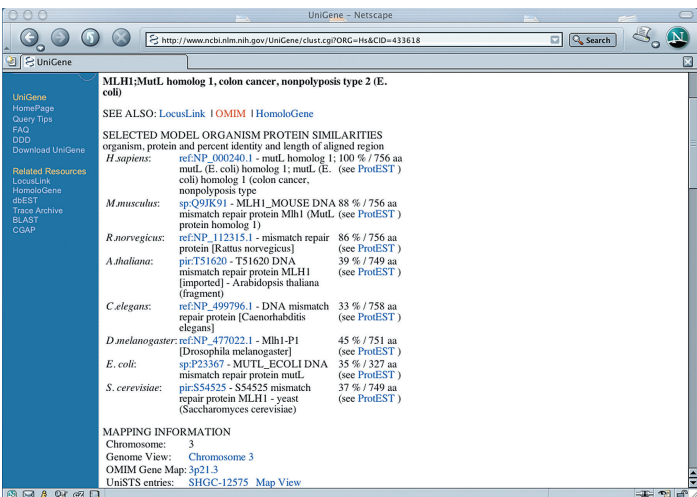


Figure 4. A UniGene display showing information on the MLH1 cluster. Each UniGene entry contains information on protein similarities, mapping data, expression information, and links to the mRNA and EST sequences comprising the cluster. See text for details.

the best match of this UniGene cluster to sequences in other organisms. Scrolling down, users find a section on mapping information; here, the gene of interest is on chromosome 3, at 3p21.3. Users can click on the *Genome View* link in this section to jump directly to the NCBI Map Viewer and look at the genomic context of the gene; the maps will be discussed in greater detail below. Following the mapping information is a rather extensive section listing what tissues the gene has been shown to be expressed in, along with a link to any available information gathered through SAGE mapping (6). Finally, the UniGene entry ends with a list of the actual mRNA and EST sequences that have been brought together to comprise this cluster.

Moving back in the list to the *H*, users can access a service at NCBI called HomoloGene. HomoloGene includes curated and calculated orthologs and homologs for genes from human, mouse, rat, and zebrafish. The curated orthologs come from the Mouse Genome Database (7), the Zebrafish Information Resource (8), and published reports in the literature. Calculated homologs and orthologs are derived from direct nucleotide sequence comparisons between all UniGene clusters from each pair of organisms. Here, the functional definition of homolog is “the best match between a UniGene cluster in one organism and a cluster in a second organism,” providing a representation of *similarity*. The functional definition of ortholog is when 2 sequences in different organisms are best matches to *one another*—a reciprocal best match—inferring that these sequences are direct descendants of a sequence in a common ancestor and most likely share common characteristics such as domain structure and biological function. Figure 5 shows the section of a HomoloGene entry for MLH1 where calculated orthologs are presented, giving the user a quick way of viewing and collecting sequence information on a given gene represented in a variety of organisms.

The final remaining alphabet block is the *V*, for variation. Clicking on the *V* brings the user to dbSNP, the database of human single nucleotide polymorphisms maintained by NCBI. The most important part of this page is the section shown in Figure 6, beginning with the header *Gene Model*. Immediately above the

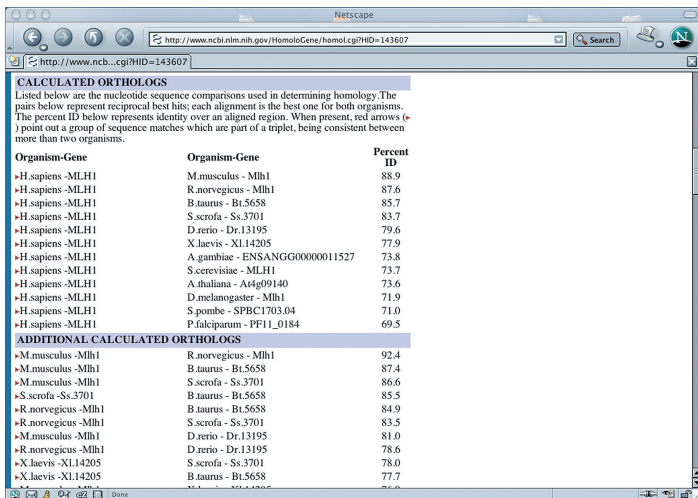


Figure 5. The HomoloGene entry for MLH1. Here, both calculated and curated orthologs to the MLH1 gene in a number of organisms are shown, as well as the percent identity between human MLH1 and its counterpart in other organisms.

table is a gray bar with a series of vertical yellow, red, and blue bars. This gray bar represents the MLH1 gene, and a key for the color-coded features is shown immediately below the gray bar to the right. Briefly, exons are shown as thicker blue bands and introns are shown as thinner blue bands. The most important vertical marks are the ones in green and red; the green marks indicate where there is an SNP that leads to a synonymous change at the protein level, whereas the red marks indicate where there is a SNP that results in a nonsynonymous change at the protein level. More information on the individual SNPs can be found by looking at the table that follows immediately below the gray bar. In this case, the 1st entry in the table shows a synonymous change (G → A, resulting in no change in the glutamine residue at position 13), whereas the 2nd

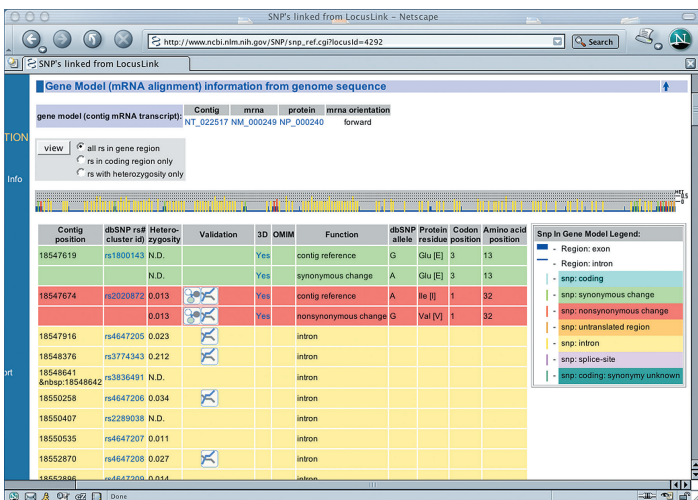


Figure 6. Single nucleotide polymorphism (SNP) data for human MLH1. The upper portion of the figure presents the gene model (position of introns and exons) and a graphical overview of where the various known SNPs for this gene are located. The table provides more detailed information about each characterized SNP. See text for details.

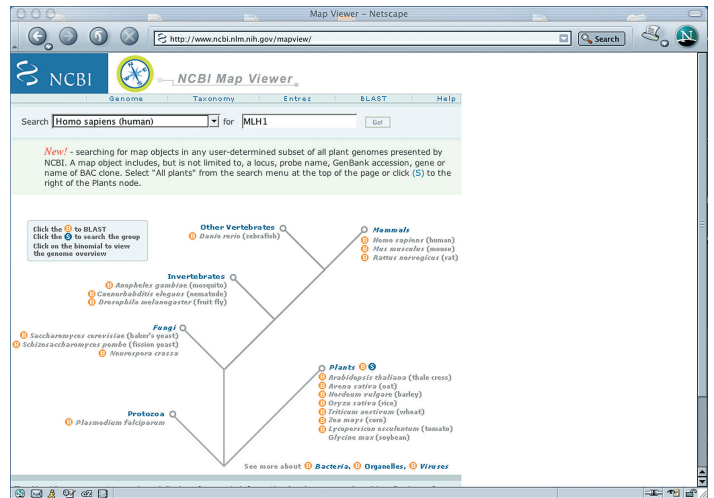


Figure 7. The NCBI Map Viewer home page. From this page, users can select any of the available organisms for which map information is available and perform targeted queries (by gene, location, or any of a number of other criteria). Information on constructing queries can be found by following the *Help* hyperlink in the upper right.

entry shows a nonsynonymous change (A → G, resulting in a change of an isoleucine to a valine at position 32). This information is extremely useful in a wide variety of studies, ranging from the more intelligent design of site-directed mutagenesis experiments to three-dimensional structural studies aimed at deciphering the net effect of a mutation on the structure (and function) of a protein.

Whereas all of these component resources have been illustrated in the context of a LocusLink search, each one of these databases can be queried individually, and the URLs for all of these resources are provided in Table 1.

GENOME BROWSERS

LocusLink obviously provides a very easy-to-use, gene-centric view of the “sequence information space,” but what if a scientist is more interested in seeing the gene of interest in context, particularly now that human genome sequencing is complete? A number of portals called *genome browsers* have been developed that allow users to access genomic data and, more importantly, view annotations that have been made on the underlying sequence data.

As alluded to above, NCBI has its own Map Viewer, a tool through which experimentally verified genes, predicted genes, genomic markers, physical maps, genetic maps, and sequence variation data can be viewed. Currently, the Map Viewer can be used to view the genomes of 19 organisms (Figure 7), with the number increasing as the genomes of more and more model organisms are sequenced. The Map Viewer is integrated into other NCBI tools, allowing one to link between the Map Viewer, LocusLink, and the main integrated information retrieval system at NCBI, Entrez.

Continuing with the MLH1 example, to find the genomic context of the MLH1 gene, one would simply change the pull-down menu marked *Search* on the Map viewer home page (<http://www.ncbi.nlm.nih.gov/mapview/>; see Figure 5) to *Human*, type “MLH1” in the text box, then press *Go*. The resulting screen is

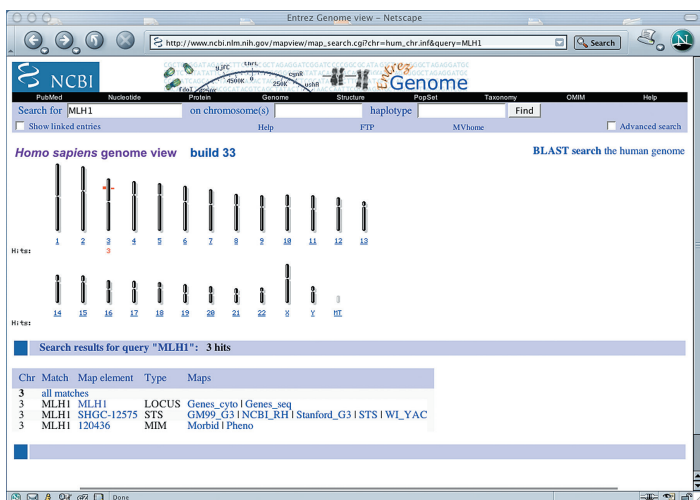


Figure 8. The results of an NCBI Map Viewer search, using “human” as the organism and “MLH1” as the query. The query returned 3 matches, one of which is to the MLH1 locus. See text for details.

shown in Figure 8. Notice that there are several red bars marking chromosome 3; below the chromosome, the number 3 appears, indicating 3 hits. Below the pictogram is a list of all the matches found for MLH1: the 1st corresponds to the gene itself (“locus”), an STS marker is next, followed by an entry from OMIM (“MIM”). Clicking on the hyperlinked *MLH1* in the “locus” line returns the map shown in Figure 9, which is the default map display. The header above the actual map gives some overview information about the map itself: there are 1906 genes on chromosome 3, and the region that is currently being displayed (the “sequence coordinates”) are from 36805K to 36948K. These numbers are also indicated in the blue bar to the left of the maps; the region displayed is also indicated by the red tick mark next to the ideogram in the blue sidebar, relative to the known cytogenetic banding patterns on chromosome 3.

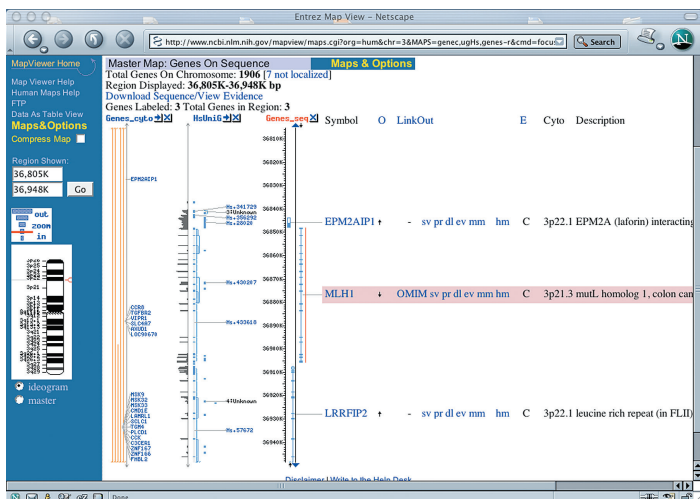


Figure 9. The default map view. Three maps are displayed: the cytogenetic gene map, the UniGene cluster map, and the “Genes_seq” map (known and putative genes that have been placed as a result of alignments of mRNAs to individual contigs). See text for details.

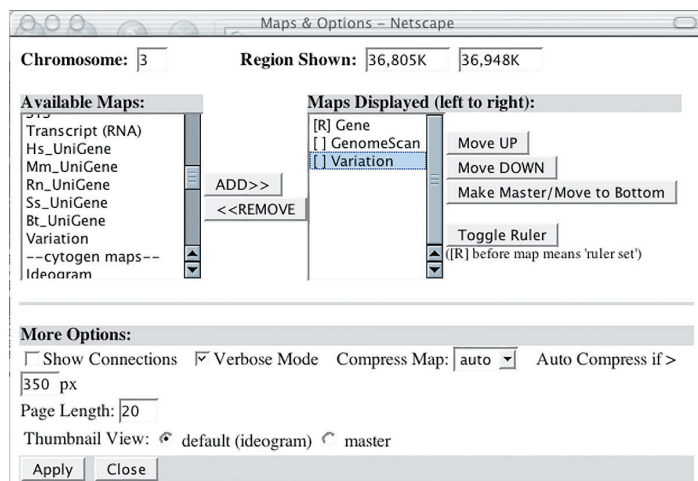


Figure 10. The Maps & Options window. This control panel is used to select from the available maps and change the order in which the maps are displayed.

Three maps are shown in the main window, to the left, as long, vertical bars. The map marked *Genes_cyto* (for “genes-cytogenetic”) shows the cytogenetic locations of genes as reported in LocusLink. Twenty genes, in addition to MLH1, have been cytogenetically mapped to this region of chromosome 3. The next map, marked *HsUniG* (for “Human UniGene”) shows the positions of UniGene clusters (described above); put another way, mRNA and EST sequences that comprise a UniGene cluster map to this region. On the left side of this particular map are gray bars that form what appears to be a histogram. These bars are intended to illustrate the density of aligned mRNAs and ESTs in this region.

The thick blue lines to the right of this map are intended to illustrate exons. The final, right-most map is labeled *Genes_seq* (for “gene sequence”). The map occupying the right-most position in any view is called the “master map,” and the information appearing to the right of all the maps pertains to that master map. Three genes are plotted on the master map in this particular view: an EPM2A-interacting protein 1, then the MLH1 gene, which was the basis of the query (highlighted in red), and finally a leucine-rich repeat interacting protein 2 (LRRFIP2). For each gene, an indication of the gene’s structure is given by the blue line running along the right side of the map, with exons being represented as thick blue bars and introns being represented as the thinner, intervening blue lines. Finally, note the arrow immediately to the right of each gene name; this arrow represents the direction of transcription for each gene.

Whereas the default map view is useful to gain a sense of what a particular genomic region looks like, there are additional maps available that may shed more light on the biological properties of a particular area of interest. To change the maps that are shown in any particular view, the user can click on the link marked *Maps & Options* that appears at the top of any Map Viewer page (see Figure 9, top). Clicking on the link brings up the Maps and Options window, allowing the user to now customize the view (Figure 10). For purposes of this example, we will remove the cytogenetic and UniGene map; to do so, the user would highlight each of these in the *Maps Displayed* list on the

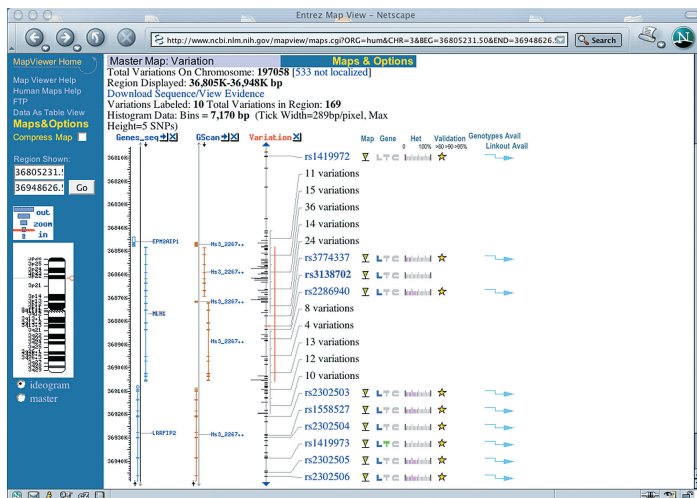


Figure 11. A new map view for the MLH1 gene, using the options shown in Figure 10. Notice that the Variation map showing all known SNPs is now the “master map.” See text for details.

right, then click <<Remove. Suppose that we wished to add the GenomeScan map (to see where all predicted genes are located) and the Variation map (to see the position of all known SNPs). To do so, select each of these from the list on the left, then click *Add>>*. When done, the window should be similar to that shown in Figure 10. Once these selections are made, the user would click the *Apply* button, and then the *Close* button. This will then recast the map as shown in Figure 11. Looking first at the 2 maps to the left, we see that the Genes_seq map and the Gscan (GenomeScan) map are not the same. The reason for this is that the GenomeScan map gives the results of a gene prediction algorithm (9), and the Genes_seq map’s annotation is based on known and putative genes that have been placed as a result of alignments of mRNAs to individual contigs. Note that the MLH1 gene is marked in red (as before, the center-most gene on the map).

The master map (the right-most map) is now the Variation map, giving a different display than before. As with the UniGene map in Figure 10, the gray bars shown to the left of the Variation map indicate the density of SNPs at any given position. Some positions are simply marked with the number of variations (for example, “11 variations”), indicating that the map is too dense to display information on each individual SNP; simply zoom out to get more information at those positions (see below). In this view, numerous SNPs can be seen, each marked with an “rs” number. Clicking on that rs number would bring the user to the dbSNP page for that particular SNP, which is similar in appearance (but not identical) to the Variation page shown in the LocusLink example above (see Figure 5). Moving across from the rs number is a series of columns of interest. The column labeled *Map* indicates whether a particular SNP has been mapped to the genome. If the SNP has been mapped to a single position, a single green down-arrow would be shown (as in Figure 11); if the SNP has been mapped to multiple positions, a double down-arrow would be shown. The column labeled *Gene* indicates whether the SNP of interest is associated with a particular genomic feature. In each row of the Gene column, notice that there is an L, T, and C either

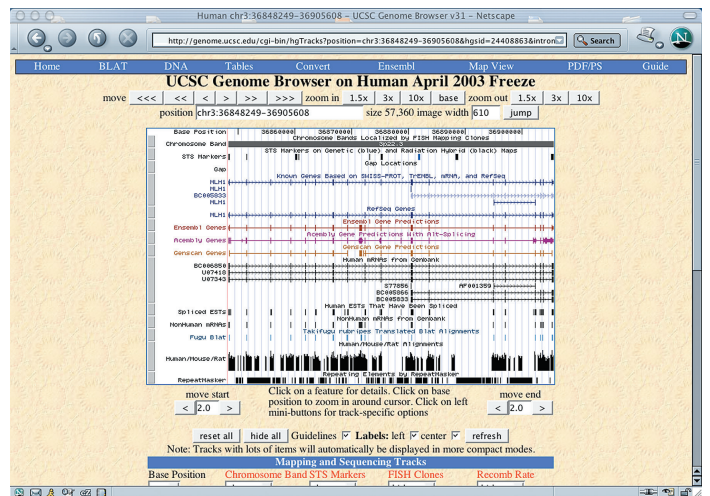


Figure 12. The UCSC Genome Browser. The region shown is for the human MLH1 gene. The overall organization is as a series of “tracks” that go from left to right, as opposed to the NCBI maps that go from top to bottom. The appearance of each track can be customized so that information appears at various densities; individual tracks can be selected or de-selected using toggles that appear below the graphic. Controls at the top of the window can be used to either zoom in or out or navigate 5’ or 3’ of the featured area.

“lit up” or “grayed out.” If the L (“locus,” blue) is lit, as it is for most of the SNPs in Figure 11, that indicates that the SNP lies within 2 kb of the 5’ end of a gene or within 500 bases of the 3’ end of a gene. If the T (“transcript,” green) is lit, the marker overlaps with a known mRNA. Finally, if the C (“coding,” orange) is lit, part or all of the SNP marker position overlaps with the coding region of a gene. The columns that follow provide additional information about the quality of the SNP marker, and more information on each of these can be found by clicking on the blue column headers.

In addition to changing the maps shown in any given view, the user can navigate by clicking anywhere on the ideogram on the left, or zoom in and out by clicking on the “out-zoom-in” picture above the ideogram. There are also short, gray bars at the top and bottom of each map that allow the user to “scroll up” or “scroll down,” moving to the next genomic segment.

This particular example only scratches the surface of what can be done with the NCBI Map Viewer. To further complicate matters, there are 2 other genome browsers that have found widespread usage and should be in the arsenal of every molecular biologist. These browsers take slightly different approaches to visualizing genomic data, and users may prefer using 1 browser to another to answer a particular biological question. The 1st of these, the UCSC Genome Browser (10), is based on the concept of “tracks,” where each track represents a particular type of annotation; this roughly corresponds to NCBI’s maps. The annotation tracks available through UCSC include known genes, predicted genes, EST alignments, and cross-species homologies, to name a few. One of the strengths of the UCSC browser lies in its ease of navigation and the ability for individual users to display their own custom annotation data on a map, allowing them to correlate their own experimental data to publicly available data. An

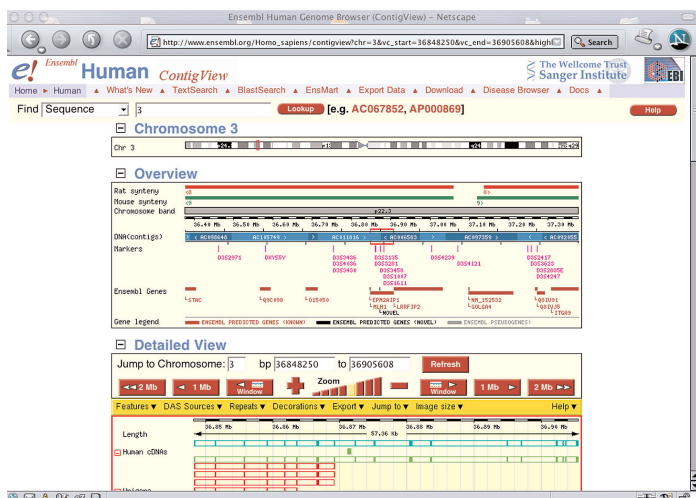


Figure 13. The Ensembl browser. The region shown is for the human MLH1 gene. The page begins with a chromosomal view, and then moves through various levels of detail. Controls in the section marked *Detailed View* can be used to either zoom in or out or navigate 5' or 3' of the featured area.

example of the UCSC display is given in Figure 12, showing the region around the MLH1 gene. Unlike the NCBI maps that run from top to bottom, the UCSC tracks run from left to right, allowing for more data to be displayed in a more intuitive fashion.

Finally, the Ensembl browser, developed by the Wellcome Trust Sanger Institute and EMBL's European Bioinformatics Institute, contains comprehensive genome annotation resulting from gene predictions and experimental data for 9 species (11). One of Ensembl's features is the ability to easily perform comparative analyses between species through the availability of DNA-DNA alignments, orthologous protein information, and large-scale synteny information. Ensembl's "ContigView" around the MLH1 gene is shown in Figure 13. As the user moves down the Web page, more detailed information is provided, moving from the

chromosomal level in the 1st panel to an overview of the region of interest in the 2nd panel to a detailed view in the 3rd panel; scrolling to the bottom of the page (not shown) would eventually bring the user to the base pair level of detail.

BEYOND THE BASICS

While space obviously precludes an in-depth treatment of any of the 3 major genome browsers, a number of useful guides and papers have been published to help biologists make intelligent use of these powerful tools. Recently, National Human Genome Research Institute published *A User's Guide to the Human Genome* (12). The majority of the guide is devoted to a series of worked examples, providing an overview of the types of data available, details on how these data can be browsed, and step-by-step instructions and strategies for using many of the most commonly used tools for sequence-based discovery. In addition, each browser's Web site provides instructional information intended to assist the novice user in using the browsers to their best advantage. The URLs for these resources are given in Table 1.

Obviously, the range of publicly available data goes well beyond just the types of data discussed in this review. Because major public sequence databases like GenBank need to be able to store data in a generalized fashion, they often do not contain more specialized types of information that would be of interest to specific groups within the biological community. Many smaller, specialized databases have emerged to fill this gap, often developed and curated by biologists "in the trenches" to address the needs of their fellow investigators. These databases, which contain information ranging from strain crosses to gene expression data, provide a valuable supplement to the major sequence repositories, and the reader is encouraged to make intelligent use of both types of databases in their searches. An annotated list of such databases can be found in the yearly database issue of *Nucleic Acids Research* (13).

Table 1. Web-based resources for sequence analysis

Major Public Sequence Databases

GenBank

<http://www.ncbi.nlm.nih.gov>

EMBL

<http://www.ebi.ac.uk/embl/index.html>

DNA Databank of Japan (DDBJ)

<http://www.ddbj.nig.ac.jp>

Gene-Centric Information Retrieval

LocusLink

<http://www.ncbi.nlm.nih.gov/LocusLink>

PubMed

<http://www.ncbi.nlm.nih.gov/Entrez>

OMIM

<http://www.ncbi.nlm.nih.gov/Omim>

UniGene

<http://www.ncbi.nlm.nih.gov/UniGene>

HomoloGene

<http://www.ncbi.nlm.nih.gov/HomoloGene>

dbSNP

<http://www.ncbi.nlm.nih.gov/SNP>

Genome Browsers

Ensembl

<http://www.ensembl.org>

NCBI Map Viewer

http://www.ncbi.nlm.nih.gov/cgi-bin/Entrez/map_search

UCSC Genome Browser

<http://genome.ucsc.edu>

Educational Resources

A User's Guide to the Human Genome

<http://www.nature.com/genomics>

Current Topics in Genome Analysis

<http://www.genome.gov/COURSE2003>

As is undoubtedly apparent by this point, there is no substitute for actually placing one's hands on the keyboard to learn how to effectively search and use genomic sequence data. Readers are strongly encouraged to take advantage of the resources presented in this review, grow in confidence and capability by working with the available tools, and begin to apply bioinformatic methods and strategies toward advancing their own research interests.

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Submitted August 27, 2003; accepted for publication October 1, 2003.

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