MINI-REVIEW

Targeting microbial pathogenic mechanisms as a novel therapeutic strategy in IBD

Paul F. Miller^{1*} \bullet

Abstract

Background Current therapy for patients suffering from inflammatory bowel diseases (IBD) is focused on inflammatory mechanisms exclusively and not the dysbiotic microbiota, despite growing evidence implicating a role for intestinal microbes in disease.

Main body Ongoing research into the intestinal microbiota of IBD patients, using new technologies and/or deeper application of existing ones, has identifed a number of microorganisms whose properties and behaviors warrant consideration as causative factors in disease. Such studies have implicated both bacteria and fungi in the pathogenesis of disease. Some of these organisms manifest mechanisms that should be amenable to therapeutic intervention via either conventional or novel drug discovery platforms. Of particular note is a deeper characterization of microbial derived proteases and their destructive potential.

Conclusion Given the steady progress on the mechanistic role of the microbiota in infammatory diseases, it is reasonable to anticipate a future in which therapeutics targeting microbial derived pathogenic factors play an important role in improving the lives of IBD patients.

Keywords Pathobiont, Dysbiosis, Pathogenesis, IBD, Microbiota, Mycobiota

Background

It is now well-accepted that the host's intestinal microbiota is a contributing factor in infammatory bowel diseases (IBD). Changes in the diversity and functionality of the gut microbial community in IBD patients compared with healthy individuals, a situation referred to as dysbiosis, has been noted for many years (Ott et al. [2004](#page-7-0), Tamboli et al. [2004\)](#page-8-0). Further defned in disease association studies as a reduction in microbial diversity (Frank et al. [2007](#page-6-0); Sartor [2008](#page-7-1)), subsequent work has sought to defne mechanisms that explain how these changes contribute to a patient's disease status and whether they are simply correlative or

Paul F. Miller

paulmiller122@gmail.com

indeed causative (Gevers et al. 2014). This distinction has important therapeutic consequences, as current IBD therapies are almost exclusively focused on the dysregulated immune system that is a hallmark of disease. Logically, immune suppressive agents have been a mainstay in disease treatment, beginning with broadly acting, non-specifc drugs such as steroids and evolving to include newer therapeutics that target specifc pathogenic inflammatory factors. This latter category includes the biologics that neutralize or sequester proinflammatory cytokines or trafficking molecules that direct immune cells to the intestinal epithelium. Unfortunately, these agents typically show efficacy in inducing clinical remission in less than half of the studied patients in pivotal clinical trials, with the responder population challenging to identify prospectively. For example, Paramsothy and colleagues reviewed registration trial data for new agents in Crohn's disease (CD) and ulcerative colitis (UC) patients, and the summary

© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit [http://creativecommons.org/licenses/by/4.0/.](http://creativecommons.org/licenses/by/4.0/)

^{*}Correspondence:

¹ Lighthouse Biopharma Consulting, LLC, 39 Emerald Glen Lane, Salem, CT 06420, USA

tables therein are informative (Paramsothy et al. [2018\)](#page-7-2). Surgery, while delayed, remains an eventual outcome for many patients (Bernstein et al. [2012](#page-6-2), Oresland and Faerden [2015](#page-7-3)).

Targeting infammation, while efective in many patients, may not address mechanisms operating at the initial phases of disease onset. Interestingly, while a large number of host susceptibility genes have been associated with IBD, several are involved in the recognition of, or response to, microbes or microbial products (Venema et al. [2017\)](#page-8-1). Given the large number of bacterial species in an individual, along with the heterogeneous composition of microbial communities across patients, the prospect of identifying individual causative organisms is daunting. Nonetheless, the feld of microbiome research has advanced steadily over the past 15 years, progressing from its initial phase of microbial cataloging to mechanistic characterization at the genetic and metabolic level (Britton and Faith 2021). The initial descriptive community membership studies, including those comparing healthy individuals with patients burdened by varying diseases, may have had the unintended efect of shifting focus away from individual species of bacteria since obvious pathogenic fngerprints didn't immediately emerge. One cause of this situation was the use of 16S sequencing to characterize microbial community composition. While quite useful and state-of-the art at the time, these methods mostly identify organisms at the family or genus level, and rarely as species. Importantly, bacterial species consist of numerous strains that can be distinguished by the presence, absence or reorganization of distinct genes that alter the functions of the organism in diferent environments. Consequently, the use of deep sequencing methods that allow strain-level characterization of organisms, particularly when coupled with culturing of these organisms from the same patients, has enabled the implication of specifc pathobionts in disease and merited their further investigation as therapeutic targets.

To further assist in bridging the gap between organism cataloging and the mechanistic implication of specifc microbes, new technologies have been developed to identify those strains that can induce disease-relevant efects in pre-clinical models, beginning with patient-derived biosamples (Palm et al. [2014;](#page-7-4) Geva-Zatorsky et al. [2017](#page-6-4); Britton et al. [2020](#page-6-5)). These tools, and several others mentioned below, offer insights into the interface between the intestinal microbiota and the host's immune system, and have provided additional means to explore the role of specifc microbes as initiators or exacerbators of disease. This review will highlight several examples of these associations, with a focus on those organisms where a greater burden of evidence exists, including identifed mechanisms of pathogenesis.

Main text

Supporting evidence for a causative role of the microbiome in IBD Before proceeding to a discussion of individual organisms, it is perhaps useful to briefy summarize other related treatment approaches that support a causal role for the microbiota in IBD. Antibiotic treatment is one such paradigm, and has been advocated as a means to alter the composition of a dysbiotic microbial community and reduce the numbers of presumed problematic pathogens, including enteric Gram-negative species such as *E. coli*. It is hoped that such treatment would also provide an opportunity for beneficial organisms to expand and restore a healthier community structure. Concerns with this approach include side efects of antibiotic use, an increased risk of *C. difficile* infection, a mismatch between the antibiotic regimen applied and the actual offending organisms in a given patient, and the emergence of antibiotic resistance (Nitzan et al. [2016\)](#page-7-5). It is challenging to summarize the overall beneft of antibiotic therapy in the treatment of IBD, as many studies are small in size and use diferent drug regimens and treatment courses. Nitzan and colleagues summarized available data from antibiotic treatment studies in CD and UC patients, noting that a greater amount of data was available for CD than for UC. In CD patients, there was a trend towards a beneft of antibiotic treatment relative to placebo, with metronidazole showing improved performance compared with ciprofloxacin in inducing clinical remission. Results from trials involving UC patients were more mixed, but overall, the analyses have shown an increased likelihood of achieving clinical remission in antibiotic treated cohorts (Nitzan et al. [2016](#page-7-5)). Thus, while the side efects and undesirable consequences associated with antibiotic treatment limit its usage, the benefts are consistent with an active role of the microbiota in IBD.

Following the same logic, correction of dysbiosis in IBD patients as an approach to disease treatment could be accomplished through fecal microbiota transplantation (FMT), which involves the instillation of intestinal bacteria in the form of a stool sample from a healthy donor to a recipient subject. This method has proven successful in the prevention of recurrent *Clostridioides difcile* infection, motivating its exploration in IBD. Imdad and colleagues reviewed a small number of placebo-controlled FMT studies involving UC patients, and concluded that while the procedure appeared to produce a roughly twofold increase in remission rates, the studies were too small to draw clear conclusions (Imdad et al. [2018\)](#page-7-6). Similarly, Fehily reviewed ffteen published FMT studies in CD patients with encouraging preliminary results. However, the authors suggest the need for large, controlled studies to solidify these fndings (Fehily et al. [2021](#page-6-6)). More recently, Feng and colleagues scrutinized published trials

of FMT for the treatment of UC. After focusing on 13 studies with the highest data quality, they concluded that FMT has a clear beneft for UC patients in inducing clinical and endoscopic remission (Feng et al. [2023\)](#page-6-7).

Against this backdrop, an alternative to the restoration of a "healthy microbiota" is the identifcation of problematic organisms in patients' dysbiotic communities, and developing therapeutic approaches based on the pathogenic mechanisms elaborated by these species. Such an approach is not without its challenges, including the translational validity of rodent models in replicating human disease as well as the technical hurdle of targeting individual organisms without afecting other benign or benefcial members of the ecosystem. Indeed, the selective removal of individual species or strains from a complex community is a daunting challenge. Currently available antibiotics would afect additional bystander organisms beyond the targeted pathogenic strains, as mentioned above. Accordingly, bacteriophages, or phages, which are viruses that infect specifc species of bacteria, have drawn attention as potential therapeutics that could be used to precisely remove organisms of interest in a targeted fashion. The intestinal microbiota is in actuality a milieu of both microbes and their infecting phages, and consequently stool samples or human waste streams can be sampled for viruses capable of killing a bacterium of interest. As bacterial resistance to infection by a specifc phage can emerge quickly, therapeutic approaches include cocktails of these viruses as well as engineered versions to increase bacterial killing (Hsu et al. [2019,](#page-7-7) Kilcher and Loasner [2019](#page-7-8)). While conceptually intriguing and promising, this approach is still under development. Finally, numerous animal models exist to support the development of drug discovery projects (Kiesler et al. [2015](#page-7-9)). Diferent models have strengths and weaknesses in supporting IBD research, and it is unclear as to which of these will be the most translationally useful in supporting the development of microbiome-targeted therapeutics. Nonetheless, substantial progress has been made in identifying candidate causative organisms in IBD as well as in progressing novel therapeutics against these, as is discussed in the following sections.

Adherent-invasive Escherichia coli (AIEC) Numerous studies have identifed an increase in the abundance of Enterobacteriaciae in the intestinal microbiota of IBD patients, including *E. coli*. A subset of *E. coli* strains was subsequently implicated in disease due to their association with, and invasion of, the intestinal epithelium. Originally implicated in IBD over 25 years ago (Darfeuille-Michaud et al. [1998](#page-6-8)), the identifcation of these strains remains challenging due to an absence of specifc virulence factors or genetically distinguishing markers (Nash et al. [2010\)](#page-7-10). Consequently, the identifcation of AIEC strains has been based on in vitro, cell-based assessments including the invasion and replication in J774 mouse macrophages as well as human epithelial Caco2 cells (Glasser et al. [2001](#page-6-9), Boudeau et al. [1999](#page-6-10)). More recently, Kittana et al. reported on the evaluation of a more comprehensive set of AIEC isolates to assess the utility of in vitro assays (Kittana et al. [2023\)](#page-7-11). These investigators systematically analyzed a collection of intestinally derived *E. coli* strains from patients and examined the relationship between in vitro and in vivo phenotypes. The ability of strains to survive and replicate in J774 macrophages and Caco2 cells in vitro was most predictive of induction of infammation in vivo. Consistent with previous observations, however, the confrmed AIEC isolates were not associated with specifc distinguishing genetic markers or known pathogenicity factors. The authors conclude that AIEC character is consistent with the adaptive behavior of *E. coli* as a species, with environmental factors selecting for favorable combinations of genes that support growth in an infammatory environment, and without the emergence of specifc, defnable clones.

Interest in AIEC as a potential driver of disease in a subset of IBD patients has motivated the search for targeted therapeutics. One approach is the development of a bacteriophage cocktail that specifcally targets these organisms. Titecat and colleagues recently described the pre-clinical safety and efficacy of a seven-phage combination therapy, called EcoActive, in a DSS mouse model of intestinal infammation (Titecat et al. [2022\)](#page-8-2). Using a collection of 210 AIEC strains from the US and Europe, this cocktail was found to be efective in lysing 95% of these isolates in vitro. High dose treatment of DSS-treated mice colonized with the AIEC strain LF82SK for 15 days resulted in a signifcant decrease in bacterial counts along with a decrease in intestinal infammatory measures. Intralytix, a co-discoverer of the EcoActive phage cocktail, announced in August 2023 that a Phase 1/2a human clinical trial had been initiated to explore the safety and efficacy of the therapeutic in patients with inactive Crohn's disease (Intralytix company website).

Taking a diferent approach, investigators at the biotech company Enterome have identifed a novel, oral, small molecule therapeutic that targets the bacterial adhesion protein FimH, used by AIEC to bind to epithelial cells. The compound, called sibofimloc (subsequently licensed to Takeda Pharmaceuticals as TAK-018), blocked the ability of AIEC to adhere to human intestinal explants and induce inflammation (Chevalier et al. [2021\)](#page-6-11). The company successfully completed a Phase 1b study in patients with active Crohn's disease, demonstrating that sibofmloc was safe and well tolerated. A Phase 2 trial in post-operative Crohn's disease patients was announced in late 2021; however, in early 2023 Takeda communicated

in a corporate portfolio update that the clinical program had been discontinued due to enrollment challenges. Consequently, the clinical hypothesis behind the FimH blocking approach remains unanswered.

Klebsiella pneumoniae This member of the Enterobacteriaciae has attracted attention more recently as a candidate pathobiont in IBD. A high-powered group of microbiome researchers used gnotobiotic mouse models to investigate the role of oral bacteria in the induction of intestinal infammation following ectopic colonization at that site (Atarashi et al. [2017](#page-6-12)). Previous studies had identifed an increased presence of oral microbes in the gut microbiota of IBD patients (Gevers et al. [2014](#page-6-1)), motivating the search for individual organisms that may be playing proinfammatory roles. Characterizing the intestinal microbiota from mice colonized by salivary bacteria from an IBD patient that had developed intestinal infammation, *K. pneumoniae* was identifed as a key driver of colitis. A related species, *K. aeromobilis*, also induced infammation in susceptible mouse models. *Klebsiella* species were also found to be enriched in alcoholic patients as well as individuals sufering from GERD and PSC. Regarding the latter disease, Nakamoto and colleagues demonstrated that the microbiota from PSC patients could induce a TH17 response in the liver of gnotobiotic mice, and that *K. pneumoniae* from these communities promoted epithelial damage and was associated with bacterial translocation in preclinical models (Nakamoto et al. [2019\)](#page-7-12).

Similar to AIEC, a therapeutic bacteriophage approach is being advanced by the biotech company BiomX as a candidate treatment for this organism. This therapeutic cocktail stemmed from a collaborative investigation on the colitigenic potential of *K. pneumoniae* strains from IBD patients that extended the fndings of Atarashi (Federici et al. [2022\)](#page-6-13). An assemblage of 5 phages was identifed that could suppress intestinal infammation in susceptible mice colonized with these strains. A healthy volunteer study demonstrated safety and tolerability of the phage cocktail, as well as the successful transit of the orally administered therapeutic to the lower GI tract. However, this candidate therapeutic is not currently listed in Biomx' active portfolio, and its status is unclear. In a similar fashion, Ichikawa and colleagues developed a phage cocktail that was efective in suppressing hepatobiliary injury in *K. pneumoniae* colonized mice (Ichikawa et al. [2023\)](#page-7-13).

Enterotoxigenic Bacteroides fragilis (ETBF) Among the better studied organisms highlighted in this review, strains of *B. fragilis* bearing the *bft* enterotoxin gene have emerged in several diferent pathogenic contexts including diarrheal disease, infammatory bowel disease and colorectal cancer (Sears [2009](#page-7-14); Valguarana 2019). This organism is able to induce a persistent colitis in standard C57BL/6 mice, making it somewhat unique among pathobionts (Rhee et al. 2009). The toxin stimulates the disruption of epithelial barriers with the concomitant release of fragments of the adherens junction protein E-cadherin (Wu et al. [2007](#page-8-3)) although E-cadherin is not the direct target. The association of this bacterial strain with IBD underscores the importance of interrogating the microbiota at greater depth than 16S sequencing, whose resolution is limited to the family or genus level. Specifcally, *B. fragilis* detection in stool samples is generally no cause for alarm; in fact, this species has been noted to have anti-infammatory properties, due in part to its capsule composition (Mazmanian et al. [2005](#page-7-16)). In stark contrast, ETBF strains containing the *bft* gene can be identifed only by deeper sequencing methods or specifc PCR detection of the toxin gene, the presence of which is sufficient to convert a common commensal into a true pathobiont (Sears [2009\)](#page-7-14).

Bft is a secreted metalloprotease that is required for the pathogenic properties associated with ETBF. It is synthesized as a pro-protein that is cleaved enzymatically to release the mature active protease. Sears and colleagues have identifed pro-infammatory and pro-oncogenic pathways induced in epithelial cells as a consequence of Bft binding. As a metalloenzyme, the prospects for identifcation and progression of specifc inhibitors as drug candidates seems promising, and among the organisms of interest in this review, the toolkit of target crystal structure (Goulas et al. [2011\)](#page-7-17), enzyme assays, cellular assays and animal models could support drug discovery eforts. Metz and colleagues described the identifcation of chenodeoxycholic acid as a candidate therapeutic for ETBF-mediated disease (Metz et al. [2019\)](#page-7-18). The molecule was shown to mitigate the cellular efects of Bft exposure through direct binding to the toxin, although an efect on protease activity was not demonstrated. Artizan Biosciences, a biotechnology company founded based on the IgA-Seq technology from the laboratory of Richard Flavell (Palm et al. [2014](#page-7-4)), has also reported on a small molecule inhibitor of the Bft protease that it is progressing towards human clinical trials (Miller [2022](#page-7-19)).

Other Bacteroides species Utilizing a sophisticated computational approach, Mills and colleagues identifed the abundance of Bacteroides-derived proteases as highly correlated with disease status in ulcerative colitis patient stool samples using an integrated multi-omics data approach (Mills et al. [2022](#page-7-20)). *B. vulgatus* proteases showed the highest correlation, with *B. dorei* also implicated. *B. vulgatus* increased epithelial barrier permeability in vitro and induced colitis in a mono-colonized IL-10 defcient mouse model; both features were reversed by administration of a broad-spectrum protease inhibitor

cocktail. Intriguingly, bacterial-derived protease activity correlated with disease severity in stool samples from this patient cohort, and high protease samples conferred higher disease scores in recipient germ free mice as compared with low protease samples obtained from patients with quiescent disease. These findings further highlight the potential role of microbial-derived proteases in IBD, as well as the power of the multi-omics strategy.

Enterococcus faecalis Among the organisms of interest as pathobionts in IBD, *E. faecalis* satisfes many criteria that qualify it for suspicion. Common commensals in many humans, the enterococci have evolved over time, notably in the context of the antibiotic era (Gilmore et al. [2013](#page-6-14)). The streamlining of these organisms' genomes has included retention of mechanisms that permit epithelial colonization and translocation. In the context of IBD, Steck and colleagues showed that *E. faecalis* is able to induce colitis in the mouse IL-10^{-/−} model, in a manner that requires expression of the GelE gelatinase (Steck et al. 2011). This secreted metalloprotease has been implicated in the ability of *E. faecalis* to translocate across the intestinal epithelial barrier, and the purifed enzyme increases epithelial barrier permeability in cell-based assays. Clinically, increased levels of *E. faecalis* were observed in IBD patients compared with healthy controls (Zhou 2016). However, GelE is produced in only a subset of *E. faecalis* strains (Galloway-Pena et al. [2011](#page-6-15)), and the presence of the *gelE* gene was not determined in the Zhou study. Related to the IBD observations with this organism, extensive work from John Alverdy and colleagues has shown that *E. faecalis* colonizes surgical anastomosis sites in patients experiencing leakage and failure of the resection. IBD patients are often the subjects of these procedures, and a role for the organism in surgical site failure was replicated in a rat intestinal resection model. Notably, the *gelE* gene was required for anastomotic failure in this model, and the mechanism involved activation of the tissue matrix metalloproteinase MMP9 (Shogan et al. [2015\)](#page-7-21).

With respect to therapeutic opportunities, the GelE metalloprotease represents a potentially druggable target, and Steck showed that GelE protease activity as well as the ability of the enzyme to disrupt epithelial barriers could be blocked by marimastat, a broadly acting MMP inhibitor (Steck et al. [2011\)](#page-8-4). These results suggest that a drug discovery effort focused on developing selective inhibitors of the bacterial enzyme may be fruitful. Alternatively, work conducted by Schnabl and colleagues showed that bacteriophage therapy targeting *Enterococcus faecalis* was efective in a mouse model of alcoholic hepatitis that was induced by this organism (Duan et al. [2019](#page-6-16)). More recently, Iida and colleagues demonstrated a putative role for *gelE-*positive *E. faecalis* in the induction of liver carcinogenesis (Iida et al. [2011](#page-7-22)). Thus, this organism appears to utilize a secreted metalloprotease in the induction of intestinal and adjacent liver diseases.

Clostridium perfringens More commonly recognized as a pathogenic cause of food-borne illness and gas gangrene, this organism has also been implicated in IBD, albeit with less literature and clinical evidence. *C. perfringens* elaborates a variety of toxins, the presence and combination of which underlie strain typing (Kiu and Hall [2018](#page-7-23)). In an examination of gut microbial sources of gelatinolytic activity, Pruteanu and colleagues identifed *C. perfringens* as the most commonly isolated producer of this protease type (Pruteanu et al. [2011](#page-7-24)). Cultured isolates contained the gene encoding the secreted metalloprotease ColA, and supernatants from these cultured strains were able to degrade the major basement membrane component type IV collagen, and also disrupted rat intestinal barrier function ex vivo using an Ussing chamber system. Subsequent work from this laboratory showed that culture supernatants could decrease the amounts of the tight junction proteins occludin and JAM-1, as well as the adherens junction protein E-cadherin. This activity appears to result from the action of both the metalloprotease ColA and the cysteine protease clostripain which were present in the culture supernatant preparations (Pruteanu and Shanahan [2013\)](#page-7-25). Intriguingly, *C. perfringens* was among the highly IgA-coated organisms identifed in IBD patient stool samples by Palm and colleagues (Palm et al. [2014\)](#page-7-4). Consequently, and given its rapid growth rate under permissive anaerobic conditions, it is tempting to speculate that a bloom of this organism in the dysbiotic microbiota of certain IBD patients could contribute to disease. As a therapeutic target, the above work from Pruteanu and colleagues showed that the epithelial disruptive activities associated with *C. perfringens* supernatants could be inhibited by the non-specifc metalloprotease inhibitor EDTA. Accordingly, it should be possible to develop optimized inhibitors of the ColA enzyme as potential drug candidates, should confdence in the causative role of this organism in disease increase. Notably, and distinct from the other metalloproteaseproducing organisms discussed here, it appears that the *colA* gene is a general feature of *C. perfringens* as a species and is conserved among all isolates studied in this context.

Candida albicans While long appreciated as a fungal opportunistic pathogen, interest in *C. albicans* as a causative agent of IBD has increased of late. Changes in the fungal component of the intestinal microbial ecosystem in the context of IBD have been noted, with increased abundance of specifc species, including *C. albicans* (Sokol et al. [2017](#page-7-26)). This organism readily colonizes mucosal surfaces and can activate pro-infammatory signaling pathways known to be induced in IBD, most notably that of IL-17 (reviewed in Ho et al. [2020\)](#page-7-27). A critical mechanistic breakthrough emerged with the discovery of the virulence factor candidalysin, a 31 amino acid peptide secreted from *C. albicans* hyphae and responsible for cellular damage induction and cytokine production (Moyes et al. 2016). The candidalysin peptide is a cleavage product from a polyprotein encoded by the *ECE1* gene, and mutants that are either lacking *ECE1* or unable to cleave the larger ECE1p protein to release the active candidalysin peptide are defective in pathogenesis (reviewed in Naglik et al. [2019\)](#page-7-29). More recently, Li et al. developed a robust fungal analysis platform utilizing IBD patient-derived samples coupled with in vitro and in vivo models to demonstrate that highly pathogenic *C. albicans* strains capable of damaging immune cells and stimulating infammatory immune responses could be isolated from the mucosa of ulcerative colitis patients (Li et al. [2022](#page-7-30)). These effects were shown to be dependent on candidalysin production. Accordingly, *C. albicans* and candidalysin bear strong consideration as potential therapeutic targets for the treatment of IBD.

Future perspectives

While not inclusive of all organisms that have garnered some measure of suspicion in the pathogenesis of IBD, the information summarized in this review is intended to shed light on a subset of strains and species where the burden of evidence is stronger. An important concept in considering individual microbes in the pathogenic process of IBD is the conditional nature of this role. From a traditional infectious diseases perspective, our microbial denizens have been distinguished as either commensals or pathogens, with an intermediate label of "opportunistic pathogens" aforded to those disease-causing organisms that are most often problematic only in individuals with underlying disease or immune insufficiency. A similar term, "pathobiont", has emerged to describe normally benign organisms that appear to be pathogenic in IBD patients (Chow et al. [2011\)](#page-6-17) or other chronic diseases (Fine et al. 2020). This can be a challenging concept, as it is unclear why such organisms would be pathogenic in one individual but seemingly innocuous in another. Host genetics is likely one factor that explains this diference.

At least 200 genetic loci have been identifed that afect the susceptibility to developing IBD (Liu 2016). Notably, a number of susceptibility genes encode functions associated with the recognition of microbial products, such as NOD2, or in the response to microbial incursion, exemplifed by the autophagy related gene ATG16L1 (Venema et al. [2017](#page-8-1)). As such, one simple explanation for the sensitivity of IBD patients to the presence of pathobionts might be that epithelial barrier disruption by these organisms results in an infux of luminal microbes that triggers a dysregulated infammatory response due to defects in the mechanisms for sensing or clearing the initial invaders. In addition, and as noted above, the abundance of several of the pathobionts highlighted here is much higher in IBD patients than in healthy carriers. It will be important to genetically characterize IBD patients who are harboring these organisms to determine if genetic factors are associated in organism-specifc ways.

The induction of disease in animal models following the introduction of these organisms, along with their increased abundance and ability to disrupt epithelial barriers, is suggestive of a role in the initiation of disease. In a clinical context, assessment of maintenance of remission is an attractive efficacy measure to include in the design of patient trials. This might merit combination studies with standard of care therapy, including biologics, to determine if efective resolution of symptoms and increased time to relapse is observed by blocking both barrier disruption by pathobionts as well as dysregulated infammation. Since several of the pathogenic factors described here are able to disrupt epithelial barriers, an increased rate of intestinal wound healing may be a beneft of blocking these virulence mechanisms. Strategies for the use of these therapies in the prevention of fares should also be explored.

The ability to identify most, if not all, of the above organisms through the use of companion diagnostic tools would create an opportunity for personalized medicine in the treatment of IBD. Examples of these technologies would include PCR assays for pathogenesis genes (e.g. *bft* in *B. fragilis*) and ELISA assays for the pathogenic factors themselves (e.g. *B. vulgatus* proteases). These could be applied to patient stool samples and would support enrollment decisions in clinical trial recruiting as well as in patient treatment paradigms if specifc therapeutics were successfully commercialized. More comprehensively, metagenomic sequencing of patient stool samples could also be used to create a more complete picture of a patient's intestinal microbiota and the community contexts in which these organisms are found. This characterization would also be valuable in following patients longitudinally to understand the impact of blocking a pathogenicity factor on the persistence and abundance of an organism.

In summary, deeper investigations into the patientspecifc behaviors of individual organisms have allowed a greater understanding of the mechanisms by which intestinal microbes can alter the epithelial environment to afect disease. Of particular note is the theme of microbial-derived intestinal proteases, the overall burden of which is higher in IBD patients than healthy individuals. Several disease-associated organisms highlighted here produce these enzymes which are mechanistically linked to their ability to cause disease in relevant models. As evidence continues to build, it is anticipated that drug developers will clinically test the microbial driver hypothesis with new therapeutics, and as a consequence bring a new level efficacy and personalized medicine to IBD patients that is complimentary to existing therapeutic approaches.

Acknowledgements

Not applicable.

Author contributions

Paul Miller conducted literature reviews and wrote the manuscript, following an invitation from the issue editor (Gerard Honig).

Funding

The author received no funding associated with the preparation of this manuscript.

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable, as Paul Miller now works independently as the sole employee of Lighthouse Biopharma Consulting

Competing interests

Paul Miller was previously an employee at Artizan Biosciences and now consults with various biopharmaceutical companies, including ones investigating the role of the microbiota in human disease.

Received: 14 December 2023 Accepted: 19 May 2024 Published online: 13 August 2024

References

- Atarashi K, Suda W, Luo C, Kawaguchi T, Motoo I, Narushima T, Kiguchi Y, Yasuma K, Watanabe E, Tanoue T, Thaiss CA, Sato M, Toyooka K, Said HS, Yamagami H, Rice SA, Gevers D, Johnson RC, Segre JA, Chen K, Kolls JK, Elinav E, Morita H, Xavier RJ, Hattori M, Honda K. Ectopic colonization of oral bacteria in the intestine drives T_H1 cell induction and inflammation. Science. 2017;358(6361):359–65. [https://doi.org/10.1126/science.aan45](https://doi.org/10.1126/science.aan4526) [26](https://doi.org/10.1126/science.aan4526).
- Bernstein CN, Loftus EV Jr, Ng SC, Lakatos BM, Epidemiology and Natural History Task Force of the International Organization for the Study of Infammatory Bowel Disease (IOIBD). Hospitalisations and surgery in Crohn's disease. Gut. 2012;61:622–9. [https://doi.org/10.1136/gutjnl-2011-301397.](https://doi.org/10.1136/gutjnl-2011-301397)
- Boudeau J, Glasser AL, Masseret E, Joly B, Darfeuille-Michaud A. Invasive ability of an *Escherichia coli* strain isolated from the ileal mucosa of a patient with Crohn's disease. Infect Immun. 1999;67:4499–509. [https://doi.org/10.](https://doi.org/10.1128/IAI.67.9.4499-4509.1999) [1128/IAI.67.9.4499-4509.1999.](https://doi.org/10.1128/IAI.67.9.4499-4509.1999)
- Britton GJ, Faith JJ. Causative microbes in host-microbiome interactions. Ann Rev Microbiol. 2021;75:223–42. [https://doi.org/10.1146/annur](https://doi.org/10.1146/annurev-micro-041321-042402) [ev-micro-041321-042402.](https://doi.org/10.1146/annurev-micro-041321-042402)
- Britton GJ, Contijoch EJ, Spindler MP, Aggarwala V, Dogan B, Bongers G, San Mateo L, Baltus A, Das A, Gevers D, Borody TJ, Kaakoush NO, Kamm MA, Mitchell H, Paramsothy S, Clemente JC, Colombel J-F, Simpson KW, Dubinsky MC, Grinspan A, Faith JJ. Defned microbiota transplant restores Th17/RORgt+ regulatory T cell balance in mice colonized with

infammatory bowel disease microbiotas. PNAS. 2020;117(35):21536–45. [https://doi.org/10.1073/pnas.1922189117.](https://doi.org/10.1073/pnas.1922189117)

- Chevalier G, Laveissi A, Desachy G, Barnich N, Sivignon A, Maresca M, Nicoletti C, Di Pasquale E, Martinez-Medina M, Simpson KW, Yajnik V, Sokol H, Plassais J, Strozzi F, Cervino A, Morra R, Bonny C. Blockage of bacterial FimH prevents mucosal infammation associated with Crohn's disease. Microbiome. 2021;9:176. <https://doi.org/10.1186/s40168-021-01135-5>.
- Chow J, Tang H, Mazmanian SK. Pathobionts of the gastrointestinal microbiota and infammatory disease. Curr Opin Immunol. 2011;23:473–80. [https://](https://doi.org/10.1016/j.coi.2011.07.010) [doi.org/10.1016/j.coi.2011.07.010.](https://doi.org/10.1016/j.coi.2011.07.010)
- Darfeuille-Michaud A, Neut C, Barnich N, Lederman E, Di Martino P, Desreumaux P, Gambiez L, Joly B, Cortot A, Colombel JF. Presence of adherent *Escherichia coli* strains in ileal mucosa of patients with Crohn's disease. Gastroenterology. 1998;115:1405–13. [https://doi.org/10.1016/s0016-](https://doi.org/10.1016/s0016-5085(98)70019-8) [5085\(98\)70019-8.](https://doi.org/10.1016/s0016-5085(98)70019-8)
- Duan Y, Llorente C, Lang S, Brandl K, Chu H, Jiang L, White RC, Clarke TH, Nguyen K, Torralba M, Shao Y, Liu J, Hernandez-Morales A, Lessor L, Rahman IR, Miyamoto Y, Ly M, Gao B, Sun W, Kiesel R, Hutmacher F, Lee S, Ventura-Cots M, Bosques-Padilla F, Verna EC, Abraldes JG, Brown RS Jr, Vargas V, Altamirano J, Caballería J, Shawcross DL, Ho SB, Louvet A, Lucey MR, Mathurin P, Garcia-Tsao G, Bataller R, Tu XM, Eckmann L, van der Donk WA, Young R, Lawley TD, Stärkel P, Pride D, Fouts DE, Schnabl B. Bacteriophage targeting of gut bacterium attenuates alcoholic liver disease. Nature. 2019;575:505–11. <https://doi.org/10.1038/s41586-019-1742-x>.
- Federici S, Kredo-Russo S, Valdes-Mas R, Kviatcovsky D, Weinstock E, Matiuhin Y, Silberberg Y, Atarashi K, Furuichi M, Oka A, Liu B, Fibelman M, Nadav Weiner I, Khabra E, Cullin N, Ben-Yishai N, Inbar D, Ben-David H, Nicenboim J, Kowalsman N, Lieb W, Kario E, Cohen T, Friedman Gefen Y, Zelcbuch L, Cohen A, Rappo U, Gahali-Sass I, Golembo M, Lev V, Dori-Bachash M, Shapiro H, Moresi C, Cuevas-Sierra A, Mohapatra G, Kern L, Zheng D, Nobs SP, Suez J, Stettner N, Harmelin A, Zak N, Puttagunta S, Bassan M, Honda K, Sokol H, Bang C, Franke A, Schramm C, Maharshak N, Sartor RB, Sorek R, Elinav E. Targeted suppression of human IBD-associated gut microbiota commensals by phage consortia for treatment of intestinal infammation. Cell. 2022;185:2879–98. [https://doi.org/10.1016/j.](https://doi.org/10.1016/j.cell.2022.07.003) [cell.2022.07.003.](https://doi.org/10.1016/j.cell.2022.07.003)
- Fehily SR, Basnayake C, Wright EK, Kamm MA. Fecal microbiota transplantation therapy in Crohn's disease: systematic review. J Gastroenterol Hepatol. 2021;36:2672–86. <https://doi.org/10.1111/jgh.15598>.
- Feng J, Chen Y, Liu Y, Lin X, Gong X, Xia R, He J, Sheng J, Cai H, Xiao C. Efficacy and safety of fecal microbiota transplantation in the treatment of ulcerative colitis: a systematic review and meta-analysis. Sci Rep. 2023;13:14494. <https://doi.org/10.1038/s41598-023-41182-6>.
- Fine RL, Vieira SM, Gilmore MS, Kriegel MA. Mechanisms and consequences of gut commensal translocation in chronic diseases. Gut Microbes. 2020;11:2170230. [https://doi.org/10.1080/19490976.2019.1629236.](https://doi.org/10.1080/19490976.2019.1629236)
- Frank DN, St Amand AL, Feldman RA, Boedeker EC, Harpaz N, Pace NR. Molecular-phylogenetic characterization of microbial community imbalances in human infammatory bowel diseases. Proc Natl Acad Sci U S A. 2007;104:13780–5. [https://doi.org/10.1073/pnas.0706625104.](https://doi.org/10.1073/pnas.0706625104)
- Galloway-Pena JR, Bourgogne A, Qin X, Murray BE. Diversity of the *fsrgelE* region of the *Enterococcus faecalis* genome but conservation in strains with partial deletions of the *fsr* operon. Appl Env Microbiol. 2011;77(2):442–51.
- Geva-Zatorsky N, Sefk E, Kua L, Pasman L, Tan TG, Ortiz-Lopez A, Yanortsang TB, Yang L, Jupp R, Mathis D, Benoist C, Kasper DL. Mining the human gut microbiota for immunomodulatory organisms. Cell. 2017;168:928–43. <https://doi.org/10.1016/j.cell.2017.01.022>.
- Gevers D, Kugathasan S, Denson LA, Vázquez-Baeza Y, Van Treuren W, Ren B, Schwager E, Knights D, Song SJ, Yassour M, Morgan XC, Kostic AD, Luo C, González A, McDonald D, Haberman Y, Walters T, Baker S, Rosh J, Stephens M, Heyman M, Markowitz J, Baldassano R, Griffiths A, Sylvester F, Mack D, Kim S, Crandall W, Hyams J, Huttenhower C, Knight R, Xavier RJ. The treatment-naïve microbiome in new-onset Crohn's disease. Cell Host Microbe. 2014;15(3):382–92. [https://doi.org/10.1016/j.chom.2014.02.005.](https://doi.org/10.1016/j.chom.2014.02.005)
- Gilmore MS, Lebreton F, van Schaik W. Genomic transition of enterococci from gut commensals to leading causes of multidrug-resistant hospital infection in the antibiotic era. Curr Opt Microbiol. 2013;16:10–6. [https://doi.](https://doi.org/10.1016/j.mib.2013.01.006) [org/10.1016/j.mib.2013.01.006.](https://doi.org/10.1016/j.mib.2013.01.006)
- Glasser AL, Boudeau J, Barnich N, Perruchot MH, Colombel JF, Darfeuille-Michaud A. Adherent invasive Escherichia coli strains from patients with

Crohn's disease survive and replicate within macrophages without inducing host cell death. Infect Immun. 2001;69:5529–37. [https://doi.org/10.](https://doi.org/10.1128/IAI.69.9.5529-5537.2001) [1128/IAI.69.9.5529-5537.2001.](https://doi.org/10.1128/IAI.69.9.5529-5537.2001)

- Goulas T, Arolas JL, Gomis-Ruth FX. Structure, function and latency of regulation of a bacterial enterotoxin potentially derived from a mammalian adamalysin/ADAM xenolog. PNAS. 2011;108:1856–61. [https://doi.org/10.](https://doi.org/10.1073/pnas.1012173108) [1073/pnas.1012173108](https://doi.org/10.1073/pnas.1012173108).
- Ho J, Camilli G, Grifths JS, Richardson JP, Kichik N, Naglik JR. *Candida albicans* and candidalysin in infammatory disorders and cancer. Immunol. 2020;162:11–6. [https://doi.org/10.1111/imm.13255.](https://doi.org/10.1111/imm.13255)
- Hsu BB, Gibson TE, Yeliseyev V, Liu Q, Lyon L, Bry L, Silver PA, Gerber GK. Dynamic modulation of the gut microbiota and metabolome by bacteriophages in a mouse model. Cell Host Microbe. 2019;25:1–12. [https://](https://doi.org/10.1016/j.chom.2019.05.001) [doi.org/10.1016/j.chom.2019.05.001.](https://doi.org/10.1016/j.chom.2019.05.001)
- Ichikawa M, Nakamoto N, Kredo-Russo S, Weinstock E, Nadav Weiner I, Khabra E, Ben-Ishai N, Inbar D, Kowalsman N, Mordoch R, Nicenboim J, Golembo M, Zak N, Jablonska J, Sberro-Livnat H, Navok S, Buchshtab N, Suzuki T, Miyamoto K, Teratani T, Fujimori S, Aoto Y, Konda M, Hayashi N, Chu P-S, Taniki N, Morikawa R, Kasuga R, Tabuchi T, Sugimoto S, Mikami Y, Shiota A, Bassan M, Kanai T. Bacteriophage therapy against pathological *Klebsiella pneumoniae* ameliorates the course of primary sclerosing cholangitis. Nat Commun. 2023;14:3261. [https://doi.org/10.1038/s41467-023-39029-9.](https://doi.org/10.1038/s41467-023-39029-9)
- Iida N, Mizukoshi E, Tamashita T, Yutani M, Seishima J, Wang Z, Arai K, Okada H, Yamashita T, Sakai Y, Masuo Y, Agustina R, Kato Y, Fujinaga Y, Oshima M, Honda M, Lebreton F, Gilmore MS, Kaneko S. Chronic liver disease enables gut *Enterococcus faecalis* colonization to promote liver carcinogenesis. Nat Cancer. 2011;2:1039–54. <https://doi.org/10.1038/s43018-021-00251-3>.
- Imdad A, Nicholson MR, Tanner-Smith EE, Zackular JP, Gomez-Duarte OG, Beaulieu DB, Acra S. Fecal transplantation for treatment of infammatory bowel disease. Cochrane Database Syst Rev Issue. 2018. [https://doi.org/](https://doi.org/10.1002/14651858.CD012774.pub2) [10.1002/14651858.CD012774.pub2](https://doi.org/10.1002/14651858.CD012774.pub2).
- Kieslier P, Fuss IJ, Strober W. Experimental models of infammatory bowel diseases. Cell Mol Gastroenterol Hepatol. 2015;1:154–70. [https://doi.org/](https://doi.org/10.1016/j.jcmgh.2015.01.006) [10.1016/j.jcmgh.2015.01.006](https://doi.org/10.1016/j.jcmgh.2015.01.006).
- Kilcher S, Loessner MJ. Engineering bacteriophages as versatile biologics. Trends Microbiol. 2019;27:355–67. [https://doi.org/10.1016/j.tim.2018.09.](https://doi.org/10.1016/j.tim.2018.09.006) [006.](https://doi.org/10.1016/j.tim.2018.09.006)
- Kittana H, Gomes-Neto JC, Heck K, Juritsch AF, Sughroue J, Xian Y, Mantz S, Segura Munoz RR, Cody LA, Schmaltz RJ, Anderson CL, Moxley RA, Hostetter JM, Fernando SC, Clarke J, Kachman SD, Cressler CE, Benson AK, Walter J, Ramer-Tait AE. Evidence for a causal role for *Escherichia coli* strains identifed as adherent-invasive in intestinal infammation. mSphere. 2023;8(2):1–16. <https://doi.org/10.1128/msphere.00478-22>.
- Kiu R, Hall LJ. An update on the human and animal enteric pathogen *Clostridium perfringens*. Emerg Microbes Inf. 2018;7:141–55. [https://doi.](https://doi.org/10.1038/s41426-018-0144-8) [org/10.1038/s41426-018-0144-8.](https://doi.org/10.1038/s41426-018-0144-8)
- Li XV, Leonardi I, Putzel GG, Semon A, Fiers WD, Kusakabe T, Lin W-Y, Gao IH, Doron I, Gutierrez-Guerrero A, DeCelie MB, Carriche GM, Mesko M, Yang C, Naglik JR, Hube B, Scherl EJ, Iliev ID. Immune regulation by fungal strain diversity in infammatory bowel disease. Nature. 2022;603:672–8. [https://](https://doi.org/10.1038/s41586-022-04502-w) [doi.org/10.1038/s41586-022-04502-w.](https://doi.org/10.1038/s41586-022-04502-w)
- Liu JZ, van Sommerer S, Huang H, Ng SC, Alberts R, Takahashi A, Ripke S, Lee JC, Jostins L, Shah T, Abedian S, Cheon JH, Cho J, Dayani NE, Franke L, Fuyuno Y, Hart A, Juyal RC, Juyal G, Kim WH, Morris AP, Poustchi H, Newman WG, Midha V, Orchard TR, Vahedi H, Sood A, Sung JY, Malekzadeh R, Westra H-J, Yamazaki K, Yang S-K, Barrett JC, Alizadeh BZ, Parkes M, Daley MJ, Kubo M, Anderson CA, Weersma RK. Association analyses identify 38 susceptibility loci for infammatory bowel disease and highlight shared genetic risk across populations. Nat Genet. 2016;47:979–86. [https://doi.](https://doi.org/10.1038/ng.3359) [org/10.1038/ng.3359](https://doi.org/10.1038/ng.3359).
- Mazmanian SK, Liu CH, Tzianabos AO, Kasper DL. An immunomodulatory molecule of symbiotic bacteria directs maturation of the host immune system. Cell. 2005;122:107–18. <https://doi.org/10.1016/j.cell.2005.05.007>.
- Metz P, Tjan MJH, Wu S, Pervaiz M, Hermans S, Shettigar A, Sears CL, Ritschel T, Dutilh BE, Boleij A. Drug discovery and repurposing inhibits a major gut pathogen-derived oncogenic toxin. Front Cell Infect Microbiol. 2019;9:364–72. [https://doi.org/10.3389/fcimb.2019.00364.](https://doi.org/10.3389/fcimb.2019.00364)
- Miller P. Identifying and addressing microbial drivers of infammatory diseases. 2022. Microbiome Movement Drug Development Conference, Boston MA.
- Mills RH, Dulai PS, Vazquez-Baeza Y, Sauceda C, Daniel N, Gerner RR, Batachari LE, Malfavon M, Zhu Q, Weldon K, Humphrey G, Carillo-Terrazas M, Goldasich LD, Bryant M, Rafatellu M, Quinn RA, Gewirtz AT, Chassaing B, Chu H, Sandborn WJ, Dorrestein PC, Knight R, Gonzalez DJ. Multi-omics analyses of the ulcerative colitis gut microbiome link *Bacteroides vulgatus* proteases with disease severity. Nat Microbiol. 2022;7:262–76. [https://doi.](https://doi.org/10.1038/s41564-021-01050-3) [org/10.1038/s41564-021-01050-3](https://doi.org/10.1038/s41564-021-01050-3).
- Moyes DL, Wilson D, Richardson JP, Mogavero S, Tang SX, Wernecke J, Hofs S, Gratacap RL, Robbins J, Runglall M, Murciano C, Blagojevic M, Thavaraj S, Kurzai O, Luo T, Kruger T, Kniemeyer O, Cota E, Bader O, Wheeler RT, Gutsmann T, Hube B, Naglik JR. Candidalysin is a fungal peptide toxin critical for mucosal infection. Nature. 2016;532:64–8. [https://doi.org/10.](https://doi.org/10.1038/nature17625) [1038/nature17625.](https://doi.org/10.1038/nature17625)
- Naglik JR, Gafen SL, Hube B. Candidalysin: discovery and function in *Candida albicans* infections. Curr Op Microbiol. 2019;52:100–9. [https://doi.org/10.](https://doi.org/10.1016/jmib.2019.06.002) [1016/jmib.2019.06.002](https://doi.org/10.1016/jmib.2019.06.002).
- Nakamoto N, Sasaki N, Aoki R, Miyamoto K, Suda W, Teratani T, Suzuki T, Koda Y, Chu P-S, Taniki N, Yamaguchi A, Kanamori M, Kamada N, Hattori M, Ashida H, Sakamoto M, Atarashi K, Narushima S, Yoshimura A, Honda K, Sato T, Kanai T. Gut pathobionts underlie intestinal barrier dysfunction and liver T helper 17 cell immune response in primary sclerosing cholangitis. Nat Microbiol. 2019;4:492–503.<https://doi.org/10.1038/s41564-018-0333-1>.
- Nash JH, Villegas A, Kropinski AM, Aguilar-Valenzuela R, Konczy P, Mascarenhas M, Ziebell K, Torres AG, Karmali MA, Coombes BK. Genome sequence of adherent-invasive *Escherichia coli* and comparative genomic analysis with other *E. coli* pathotypes. BMC Genomics. 2010;11:667. [https://doi.org/10.](https://doi.org/10.1186/1471-2164-11-667) [1186/1471-2164-11-667](https://doi.org/10.1186/1471-2164-11-667).
- Nitzin O, Elias M, Peretz A, Saliba W. Role of antibiotics for treatment of infammatory bowel disease. World J Gastroenterol. 2016;22:1078–87. [https://](https://doi.org/10.3748/wjg.v22i3.1078) [doi.org/10.3748/wjg.v22i3.1078.](https://doi.org/10.3748/wjg.v22i3.1078)
- Oresland T, Faerden AE. Surgery in the age of biological treatment. Scand J Gastroent. 2015;50:121–7. [https://doi.org/10.3109/00365521.2014.](https://doi.org/10.3109/00365521.2014.972445) [972445.](https://doi.org/10.3109/00365521.2014.972445)
- Ott SJ, Musfeldt M, Wenderoth DF, Hampe J, Brant O, Fölsch UR, Timmis KN, Schreiber S. Reduction in diversity of the colonic mucosa associated bacterial microflora in patients with active inflammatory bowel disease. Gut. 2004;53:685–93. [https://doi.org/10.1136/gut.2003.025403.](https://doi.org/10.1136/gut.2003.025403)
- Palm NW, de Zoete MR, Cullen TW, Barry NA, Stefanowski J, Hao L, Degnan PH, Hu J, Peter I, Zhang W, Ruggiero E, Cho JH, Goodman AL, Flavell RA. Immunoglobulin A coating identifes colitigenic bacteria in infammatory bowel disease. Cell. 2014;158:1000–10. [https://doi.org/10.1016/j.cell.2014.](https://doi.org/10.1016/j.cell.2014.08.006) [08.006](https://doi.org/10.1016/j.cell.2014.08.006).
- Paramsothy S, Rosenstein AK, Mehandru S, Colombel J-F. The current state of the art for biological therapies and new small molecules in infammatory bowel disease. Mucosal Immunol. 2018;11:1558–70. [https://doi.org/10.](https://doi.org/10.1038/s41385-018-0050-3) [1038/s41385-018-0050-3](https://doi.org/10.1038/s41385-018-0050-3).
- Pruteanu M, Shanahan F. Digestion of epithelial tight junction proteins by the commensal *Clostridium perfringens*. Am J Physiol Gastrointest Liver Physiol. 2013;305:G470–8. [https://doi.org/10.1152/ajpgi.00316.2012.](https://doi.org/10.1152/ajpgi.00316.2012)
- Pruteanu M, Hyland NP, Clarke DJ, Kiely B, Shanahan F. Degradation of the extracellular matrix Components by bacterial-derived metalloproteases: implications for infammatory bowel diseases. Infamm Bowel Dis. 2011;17:1189–200. [https://doi.org/10.1002/ibd.21475.](https://doi.org/10.1002/ibd.21475)
- Rhee KJ, Wu S, Wu X, Huso DL, Karim B, Franco AA, Rabizadeh S, Golub JE, Mathews LE, Shin J, Sartor RB, Golenbock D, Hamad AR, Gan CM, Housseau F, Sears CL. Induction of persistent colitis by a human commensal, *Bacteroides fragilis*, in wild-type C57BL/6 mice. Infect Immun. 2009;77:1708–18.
- Sartor RB. Microbial infuences in infammatory bowel diseases. Gastroenterology. 2008;134:577–94.<https://doi.org/10.1053/j.gastro.2007.11.059>.
- Sears CL. Enterotoxigenic *Bacteroides fragilis*: a rogue among symbiotes. Clin Micro Revs. 2009;22:349–69.<https://doi.org/10.1128/CMR.00053-08>.
- Shogan BD, Belogortseva N, Luong PM, Zaborin A, Lax S, Bethel C, Ward M, Muldoon JP, Singer M, An G, Umanskiy K, Konda V, Shakhsheer B, Luo J, Klabbers R, Hancock LE, Gilbert J, Zaborina O, Alverdy JC. Collagen degradation and MMP9 activation by *Enterococcus faecalis* contribute to intestinal anastomotic leak. Sci Trans Med. 2015;7:28668. [https://doi.org/](https://doi.org/10.1126/scitranslmed.3010659) [10.1126/scitranslmed.3010659](https://doi.org/10.1126/scitranslmed.3010659).
- Sokol H, Leducq V, Aschard H, Pham H-P, Jegou S, Landman C, Cohen D, Liguori G, Bourrier A, Nion-Larmurier I, Cosnes J, Seksik P, Langella P, Skurnik

D, Richard ML, Beaugerie L. Fungal microbiota dysbiosis in IBD. Gut. 2017;66:1039–48. <https://doi.org/10.1136/gutjnl-2015-310746> .

- Steck N, Hofmann M, Sava IG, Kim SC, Hahne H, Tonkonogy SL, Mair K, Krueger D, Pruteanu M, Shanahan F, Vogelmann R, Schemann M, Kuster B, Sartor RB, Haller D. *Enterococcus faecalis* metalloprotease compromises epi thelial barrier and contributes to intestinal infammation. Gastroenterol. 2011;141:959–71. <https://doi.org/10.1053/j.gastro.2011.05.035> .
- Tamboli CP, Neut C, Desreumaux P, Colombel J-F. Dysbiosis in infammatory bowel disease. Gut. 2004;53:1–4.
- Titécat M, Rousseaux C, Dubuquoy C, Foligné B, Rahmouni O, Mahieux S, Desreumaux P, Woolston J, Sulakvelidze A, Wannerberger K, Neut C. Safety and efficacy of an AIEC-targeted bacteriophage cocktail in a mice colitis model. J Crohns Colitis. 2022;16(10):1617–27. [https://doi.org/10.](https://doi.org/10.1093/ecco-jcc/jjac064) [1093/ecco-jcc/jjac064](https://doi.org/10.1093/ecco-jcc/jjac064) .
- Valguarnera E, Wardenburg JB. Good gone bad: one toxin away from disease for *Bacteroides fragilis*. J Mol Biol. 2020;432:765–85. [https://doi.org/10.](https://doi.org/10.1016/j.jmb.2019.12.003) [1016/j.jmb.2019.12.003](https://doi.org/10.1016/j.jmb.2019.12.003) .
- Venema WTCU, Voskuil MD, Dijkstra G, Weersma RK, Festen AM. The genetic background of infammatory bowel disease: from correlation to causality. J Pathol. 2017;241:146–58. <https://doi.org/10.1002/path.4817> .
- Wu S, Rhee KJK, Zhang M, Franco A, Sears CL. *Bacteroides fragilis* toxin stimu lates intestinal epithelial cell shedding and gamma-secretase dependent E-cadherin cleavage. J Cell Sci. 2007;120:1944–52.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in pub lished maps and institutional afliations.

Paul F. Miller is an experienced pharmaceutical researcher and executive. He conducted and led anti-infective research at Warner Lambert Co., Pfzer and AstraZeneca, and then held the position of Chief Scientific Officer at Synlogic and Artizan Biosciences, two companies involved in microbiome-related research.