

Guizhi Fuling Wan as a Novel Agent for Intravesical Treatment for Bladder Cancer in a Mouse Model

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Alternative intravesical agents are required to overcome the side effects currently associated with the treatment of bladder cancer. This study used an orthotopic bladder cancer mouse model to evaluate Guizhi Fuling Wan (GFW) as an intravesical agent. The effects of GFW were compared with those of mitomycin-C (Mito-C) and bacille Calmette-Guérin (BCG). We began by evaluating the response of the mouse bladder cancer cell line MB49 to GFW treatment, with regard to cell viability, cell cycle progression and apoptosis. MB49 cells were subsequently implanted into the urothelial walls of the bladder in female C57BL/6 mice. The success of the model was confirmed by the appearance of hematuria and tumor growth in the bladder. Intravesical chemotherapy was administered in accordance with a published protocol. *In vitro* data revealed that GFW arrested MB49 cell cycle in the G0/G1 phase, resulting in the suppression of cell proliferation and induced apoptosis. One possible mechanism underlying these effects is an increase in intracellular reactive oxygen species (ROS) levels leading to the activation of ataxia telangiectasia-mutated (ATM)/checkpoint kinase 2 (CHK2) and ATM/P53 pathways, thereby mediating cell cycle progression and apoptosis, respectively. This mouse model demonstrates the effectiveness of GFW in the tumor growth, with results comparable to those achieved by using BCG and Mito-C. Furthermore, GFW was shown to cause only mild hematuria. The low toxicity of the compound was confirmed by a complete lack of lesions on bladder tissue, even after 10 consecutive treatments using high concentrations of GFW. These results demonstrate the potential of GFW for the intravesical therapy of bladder cancer.

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INTRODUCTION

Urothelial carcinoma is the second most common cause of death among genitourinary tumors (1). More than 90% of urothelial

carcinoma is transitional cell carcinoma, 20% of which develops into muscle invasive disease with a substantial risk of distant metastasis (2). Despite the fact

that urothelial carcinoma can originate in any part of the urinary tract, most cases begin in the urinary bladder (3). Bladder urothelial carcinoma is heterogeneous and can be histologically subdivided into low-grade and high-grade diseases. Low-grade urothelial carcinoma is uniformly noninvasive; however, high-grade urothelial carcinoma can be classified according to the depth of invasion into the muscle layer, as follows: nonmuscle invasive bladder cancer (Tis, Ta, T1) and muscle-invasive bladder cancer (\geq T2) (4). Patients diagnosed with nonmuscle invasive bladder cancer face a high risk of recurrence (5), which necessitates the additional use of intravesical chemotherapy or bacille Calmette-Guérin (BCG) as a supplement to transurethral resection (6,7). However, mitomycin-C (Mito-C) has been shown to induce

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cystitis and hematuria (8,9), and BCG is associated with cystitis and granulomatous inflammation (10,11). Thus, novel intravesical agents with enhanced efficacy and minimal toxicity are required for the treatment of patients with bladder cancer.

The traditional Chinese herbal formula, Guizhi Fuling Wan (GFW), has recently been shown to have anticancer effects on hepatocellular carcinoma (12,13) and cervical cancer (14). We have previously shown that GFW has a potent inhibitory effect on cell proliferation by inducing cell cycle arrest and apoptosis in human bladder cancer cell lines, BFTC 905 and TSGH 8301, with only weak cytotoxicity effects on normal human urothelial cells (15).

Recent studies have identified several traditional Chinese herbal remedies capable of inducing apoptosis in cancer cells by increasing the level of reactive oxygen species (ROS) (16–18). ROS are oxygen-containing chemically reactive molecules that can cause considerable damage to key cellular molecules, such as lipids, proteins and DNA (19,20). ROS-induced DNA damage exceeding cellular repair capacity can lead to apoptosis. Furthermore, the activation of ataxia telangiectasia-mutated (ATM) kinase signaling and tumor suppressor p53 was identified as an important cellular response to ROS-induced DNA damage (21,22). The generation of excess ROS has also been shown to initiate cell cycle arrest, which can lead to the induction of p53-dependent apoptosis (23). This study sought to identify the mechanism underlying ROS involvement in the GFW-related suppression of cell growth and induced cell death in bladder cancer. We also applied a murine orthotopic bladder tumor model to evaluate the *in vivo* antitumor effects of intravesical GFW and compare the treatment outcomes with those obtained using Mito-C or BCG.

MATERIALS AND METHODS

Preparation of Agents and Cell Culture

GFW herbal extract was purchased, validated and prepared as previously

described (15). Mito-C (Kyowa Hakko Kirin) and BCG (Sanofi Pasteur Limited) were dissolved in normal saline buffer at 1 mg/mL to provide stock solutions, which were then diluted to 0.4 mg/mL and 1×10^8 CFU/3 mL, respectively. Murine urothelial carcinoma MB49 cells (gift from Dr. Timothy L. Raliff, Purdue Cancer Center) were cultured in RPMI 1640 supplemented with 10% fetal bovine serum (FBS), 100 units/mL penicillin and 100 µg/mL streptomycin in an incubator in a humidified atmosphere containing 5% CO₂ at 37°C.

Cell Cycle Analysis

MB49 cells were seeded in six-well plates at 2×10^5 cells per well and cultured for 24 h, before being starved in medium without supplemental FBS for 24 h, followed by treatment with 0.5, 1 and 2 mg/mL GFW for 24 h. After treatment, the cells were trypsinized, washed with phosphate-buffered saline (PBS) and fixed by using methanol at 4°C overnight, followed by incubation with RNase A and propidium iodide (Sigma) for 30 min at room temperature. The cells were then analyzed by using a BD FACSCanto II flow cytometer (BD Biosciences) in conjunction with Mod Fit LT™ 3.3 software.

Apoptosis Analysis

MB49 cells were seeded in six-well plates at 2×10^5 cells per well and cultured for 24 h before being treated with 0.5, 1 or 2 mg/mL of GFW for 24 h. Cell apoptosis was detected using the Annexin V-FITC Apoptosis Detection Kit (BioVision) in accordance with the manufacturer's protocol and analyzed by BD FACSCanto II flow cytometer (BD Biosciences) in conjunction with Mod Fit LT™ 3.3 software.

Reactive Oxygen Species Detection

This assay used cell-permeable fluorogenic probe 2', 7'-dichlorodihydrofluorescein diacetate (DCFH-DA). DCFH-DA can be deacetylated by using cellular esterases to nonfluorescent DCFH, which can be rapidly oxidized to highly fluorescent

DCF by intracellular ROS. The intensity of fluorescence is proportional to the intracellular ROS levels.

Briefly, MB49 cells were seeded in 10-cm plates at 2×10^6 cells per plate and cultured for 24 h. The cells were starved in medium without supplemental FBS for 24 h before being pre-incubated with 20 µmol/L DCFH-DA for 1 h. This step was then followed by treatment with 2 mg/mL GFW (experimental group) or 50 µmol/L H₂O₂ (positive control group) for 3 h. After collecting and washing the cells with PBS, they were immediately analyzed by using BD FACSCanto II flow cytometer (BD Biosciences) in conjunction with Mod Fit LT™ 3.3 software.

Western Blot Analysis

MB49 cells were seeded in six-well plates at 2×10^5 cells per well and cultured for 24 h. The cells were starved in medium without supplemental FBS for 24 h and then treated with 2 mg/mL GFW for various durations. Total cellular proteins were extracted, and the protein concentrations in the extracts were determined by using the Bio-Rad Protein Assay reagent (Bio-Rad) with bovine serum albumin as a standard. Each cell lysate (30 µg) was separated on SDS-PAGE, transferred electrophoretically to polyvinylidene fluoride transfer membranes and probed by using antibodies—1:1,000 diluted antibody against caspase-3, ATM, CHK2, phospho-CHK2, P53, phospho-P53 and P21 (Cell Signaling). The protein band that was specifically bound to the primary antibody was detected by using the BioSpectrum 800 system (UVP) after incubating with horseradish peroxidase (HRP)-conjugated secondary antibodies for 1.5 h at room temperature.

Murine Orthotopic Bladder Tumor Model and Intravesical Therapy

Female C57BL/6 mice aged 6 wks were provided by the National Laboratory Animal Center and maintained at

our animal care facility for 2 wks before use. All experiments were approved by the Institutional Animal Care and Use Committee of National Chung Cheng University (IACUC no. 1020103).

The implantation of MB49 murine bladder cancer cells into C57BL/6 mice was performed as previously described (2) with only slight modifications. Mice were anaesthetized through the administration of 1.5% Isoflurane gas, whereupon a 24-gauge Teflon intravenous catheter (BD Biosciences) was inserted through the urethra into the bladder. In preparation for tumor implantation, 100 μ L 0.1 mol/L AgNO₃ was injected into the bladder to induce a chemical lesion on the bladder wall. After 10 s, the content was washed out through the transurethral infusion of 1 mL PBS. A suspension of $5 \times 10^5/100 \mu$ L MB49 cells was then instilled into the bladder. The mice were randomly assigned to five groups (seven mice per group). Twenty-four hours after tumor implantation, the experimental group underwent intravesical therapy with 100 μ L GFW (180 mg/mL), 100 μ L Mito-C (0.4 mg/mL) or PBS every other day, or 100 μ L BCG (1×10^8 CFU/3 mL) therapy every week (Figure 1A). At the same time, the control group received no treatment (Figure 1B). After injection of GFW, Mito-C, BCG or PBS into the mouse bladders, the external urethral orifice of each mouse was blocked for 1 h by using a small piece of Micropore surgical tape. The tape was subsequently removed, and the mice were returned to their original cages. Healthy mice that did not undergo tumor implantation were used as a control to evaluate the cytotoxic effects of GFW on the epithelium of a normal bladder. GFW treatment was administered 10 times every other day to healthy mice with or without AgNO₃-induced lesion.

In Vivo Magnetic Resonance Detection of Bladder Tumors

Murine bladder tumor cells implanted into mouse bladders (as described above) were scanned by using magnetic resonance

imaging (MRI) on d 0, 4, 7 and 11 after GFW treatment. MRI data were acquired by using a 300-MHz (7 T) Apollo NMR spectrometer (Tecmag) capable of microimaging. Phantom images were obtained by using microimaging probe heads and a 32-mm I.D. imaging RF coil (Doty Scientific) with a self-shielded gradient system and maximum strength of 1,000 mT/m in the x, y and z directions (Resonance Research). All MRI data were generated by using Spin-Echo pulse sequences. The echo time (TE) and repetition time (TR) were 15 ms and 7 s, respectively.

Statistical Analysis

Numerical data are expressed as either mean \pm standard error of the mean (SEM) or as a percentage relative to the untreated control. Differences between the experiment and control groups were determined by using one-way analysis of variance (ANOVA) followed by Dunnett test. Differences were considered significant when the *p* value was <0.05 or within the 95% confidence interval.

All supplemental materials are available online at www.molmed.org.

RESULTS

GFW Inhibits MB49 Cell Proliferation through Cell Cycle Arrest and Induced Apoptosis

In a previous study, we demonstrated the inhibitory effects of GFW on the proliferation of human bladder cancer cells with minimal toxicity to normal urothelial cells. In this study, we sought to determine the effectiveness of GFW in the treatment of mouse bladder cancer cell line MB49. The initial results of *in vitro* cell viability assays revealed that GFW suppresses the proliferation of MB49 cells in a dosage-dependent manner, presenting a half-maximal inhibitory concentration (IC₅₀) of approximately 0.95 mg/mL (Supplementary Figure S1), in accordance with its effect on human bladder cancer cell line TSGH 8301 (15).

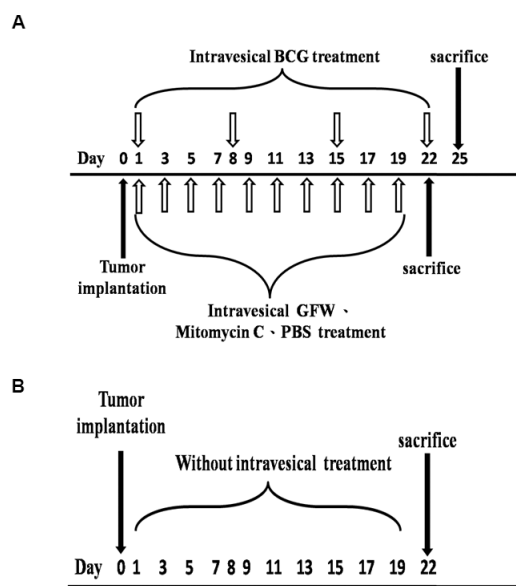


Figure 1. Strategy used in intravesical treatment in a murine orthotopic bladder tumor model. Time schedules of treatment groups (A) and nontreatment control groups (B) are shown. Twenty-four hours after tumor implantation, animals in the treatment group underwent intravesical therapy with 100 μ L GFW (180 mg/mL), 100 μ L Mito-C (0.4 mg/mL) or PBS every other day (empty arrows pointing up), or 100 μ L BCG (1×10^8 CFU/3 mL) therapy every week (empty arrows pointing down), while the control group received no treatment.

To identify the mechanism by which GFW inhibits the growth of MB49 cells, we sought to determine whether GFW treatment hinders the cell cycle progression. Flow cytometry results demonstrated that exposure to GFW at 1 and 2 mg/mL for 24 h significantly increased the percentage of MB49 cells in the G1 phase ($p < 0.01$ and $p < 0.001$, respectively; Figure 2A), while decreasing the percentage in the S phase ($p < 0.05$). These results suggest that GFW treatment leads to cell cycle arrest in the G1 phase. After administration of 1 and 2 mg/mL GFW, we observed a significant increase in the proportion of MB49 cells in the G1 phase. These values are correlated with the reduction in the proportion in the S phase at these concentrations.

We also observed that treating MB49 cells with 2 mg/mL for 24 h induced cellular apoptosis, as evidenced by the sub-G1 population, which was proportionately larger than that of the untreated control (Figure 2A). We subsequently assessed the effects of GFW treatment at concentrations of 0.5, 1 and 2 mg/mL for 24 h on the induction of apoptosis. Our results revealed a significant increase in the instances of apoptosis among cells treated with GFW at concentrations of 1 and 2 mg/mL (Figure 2B). GFW treatment at 1 and 2 mg/mL (Figure 2B) led to a proportional increase in the number of necrotic cells; however, we observed far fewer necrotic cells than apoptotic cells. We subsequently assessed the effects of GFW treatment at a concentration of 2 mg/mL over various durations (0, 24, 32 and 48 h) on the induction of caspase-3 and its cleaved form, which is a hallmark of apoptosis in MB49 cells. The results of Western blot analysis (Figure 2C) revealed a significant increase in the level of cleaved caspase-3 after GFW treatment for 48 h. In the context of the flow cytometry profile, which revealed significant increases in sub-G1 and apoptotic populations, these results demonstrate that GFW induced apoptosis in MB49 cells.

GFW Increases Intracellular ROS Levels and Activates ATM/CHK2 Pathway in the Mediation of Cell Cycle Regulatory Proteins in MB49 Cells

One promising chemotherapeutic strategy involves dramatically increasing intracellular ROS. This approach has been shown to induce random damage (24–28) and/or apoptosis (29–31) in cancer cells, with little or no effect on normal cells, because of their lower basal ROS levels, which renders them less susceptible to the adverse effects brought about by chemotherapeutic agents. To determine if GFW-mediated cell cycle arrest and apoptosis could be attributed to the generation of ROS, we treated MB49 cells with GFW for 3 h and then measured the ROS levels. As shown in Figure 3A, the mean percentages of ROS-positive cells (M2) were as follows: untreated control group, $2.1 \pm 0.7\%$; H_2O_2 treatment group, $4.2 \pm 0.1\%$; and GFW treatment group, $12.6 \pm 0.9\%$. The GFW-induced accumulation of ROS was three times higher than that induced by H_2O_2 . These results indicate that GFW enhances the intracellular accumulation of ROS in MB49 cells.

Previous studies have demonstrated that the expression of ATM induced by ROS (22,32) can mediate cell cycle checkpoints through the activation of CHK2 (33,34). Thus, we examined the induction of ATM, CHK2 and Thr68-phosphorylated CHK2 protein levels in GFW-treated MB49 cells. After treatment with GFW (2 mg/mL) for various durations, cell lysates were prepared for detection by Western blot analysis by using antibodies specific to ATM, CHK2 and Thr68-phosphorylated CHK2. As shown in Figure 3B, we observed a significant increase in ATM expression in MB49 cells after treatment for 6 h. This step was immediately followed by the activation of CHK2, as indicated by a significant increase in Thr68-phosphorylated CHK2 protein levels. The concurrent use of antibodies specific to P21, P53 and Ser15-phosphorylated P53 prompted a

significant increase in the levels of P21 and Ser15-phosphorylated P53 protein in MB49 cells, following GFW treatment for 6 h (Figure 3C). These results indicate that GFW induces the generation of ROS and the subsequent activation of ATM/CHK2 signaling leading to the induction of Ser15-phosphorylated P53 and P21.

GFW Inhibits Tumor Growth in a Murine Orthotopic Bladder Tumor Model

Our results in a previous study suggested that high concentrations of GFW *in vitro* could have cytotoxic effects on normal urothelial cells (15). In view of the fact that high concentrations of GFW were used in this *in vivo* study, we sought to determine whether GFW has cytotoxic effects on the urothelium of normal mouse bladders. Hematoxylin and eosin (H&E) staining was used to observe mouse bladder tissue after 10 consecutive GFW treatments with or without $AgNO_3$ chemical lesions. All cases revealed bladders with a normal appearance and regular urothelium. Even 10 consecutive treatments with 180 mg/mL GFW was insufficient to harm a normal mouse bladder urothelium (Figure 4A). Consequently, GFW was applied at a concentration of 180 mg/mL in all subsequent *in vivo* experiments.

Orthotopic bladder tumors were then generated in 35 mice that were randomly assigned to five groups (seven mice per group). Except for the control group (positive, Supplementary Figure S3), which showed a slight decrease in body weight from 10 d after tumor implantation, all other groups presented a steady body weight throughout the experiment (Supplementary Figure S2). After the implantation of MB49 murine bladder cancer cells for 9–11 d, hematuria was observed in all mice, regardless of whether they underwent treatment (Table 1). This result demonstrates the successful implantation of MB49 cells and subsequent growth of tumors. It is interesting to note that serious hematuria was observed

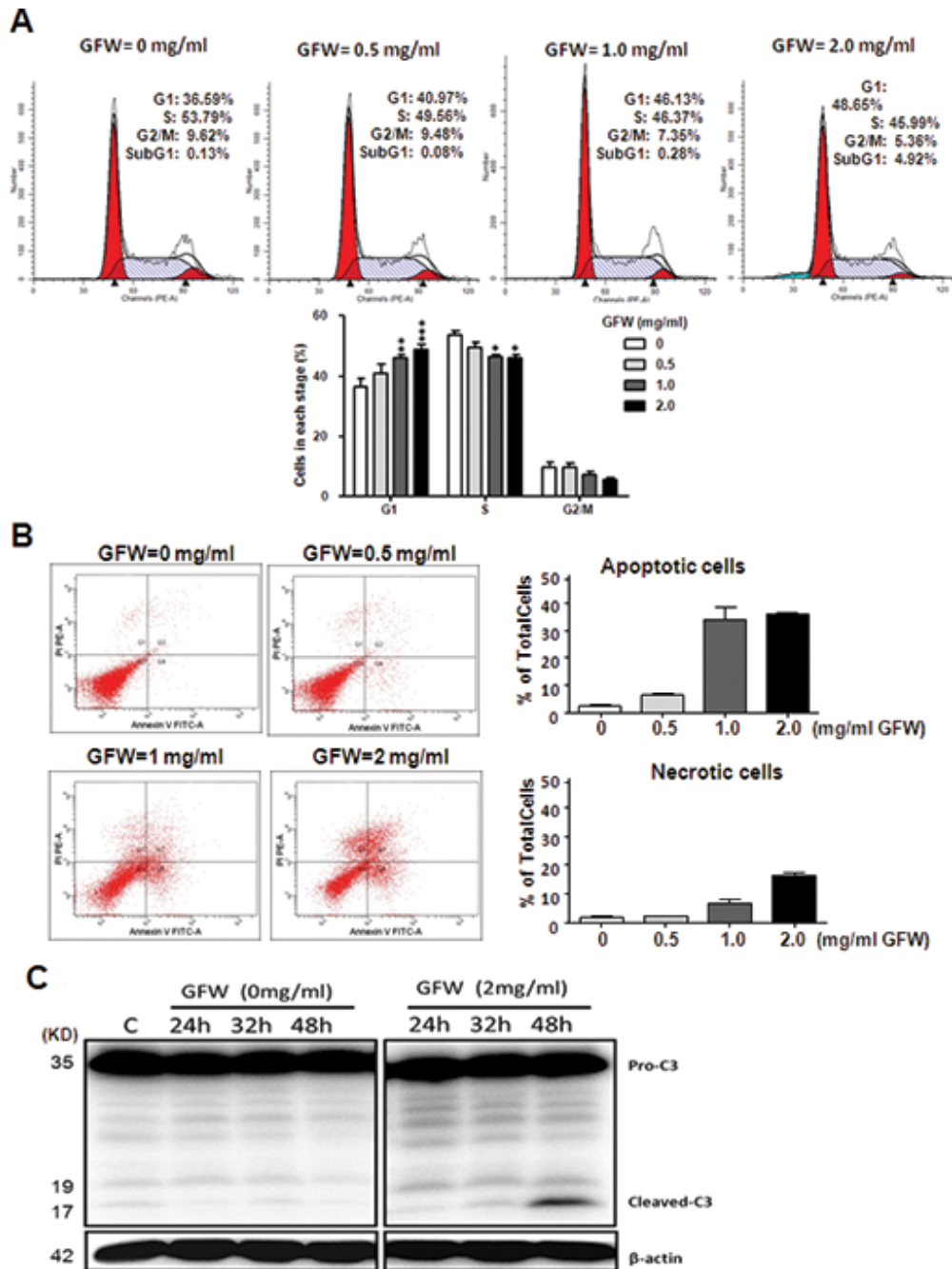


Figure 2. Effects of GFW on cell cycle and induction of apoptosis in mouse urothelial cell line, MB49. (A) MB49 cells were treated with GFW at concentrations of 0.5, 1 and 2 mg/mL for 24 h and then stained with propidium iodide and analyzed using flow cytometry. Data are represented as mean \pm SEM (n = 4). Significant differences from the control are indicated by * $p < 0.05$, ** $p < 0.01$ or *** $p < 0.001$, as determined by one-way ANOVA and Dunnett comparison test. (B) Analysis of apoptosis using flow cytometry. MB49 cells were treated with GFW at concentrations of 0.5, 1 and 2 mg/mL for 24 h, stained with propidium iodide and Annexin V and then analyzed using flow cytometry. The lower left quadrant (Q3) represents viable cells, the upper left quadrant (Q1) represents necrotic cells, the lower right quadrant (Q4) represents early apoptotic cells and the upper right quadrant (Q2) represents nonviable late apoptotic cells. The relative percentages include early and late apoptotic cells. Data are represented as mean \pm SEM of three independent experiments. Significant differences from the untreated control are indicated by * $p < 0.05$, ** $p < 0.01$ or *** $p < 0.001$, as determined by one-way ANOVA and Dunnett comparison test. (C) Analysis of caspase-3 activation in GFW-treated cells. Western blot analysis was performed to detect caspase-3 and its cleaved form in MB49 cells after treatment with 2 mg/mL GFW for various durations. β -Actin protein was used as the loading control.

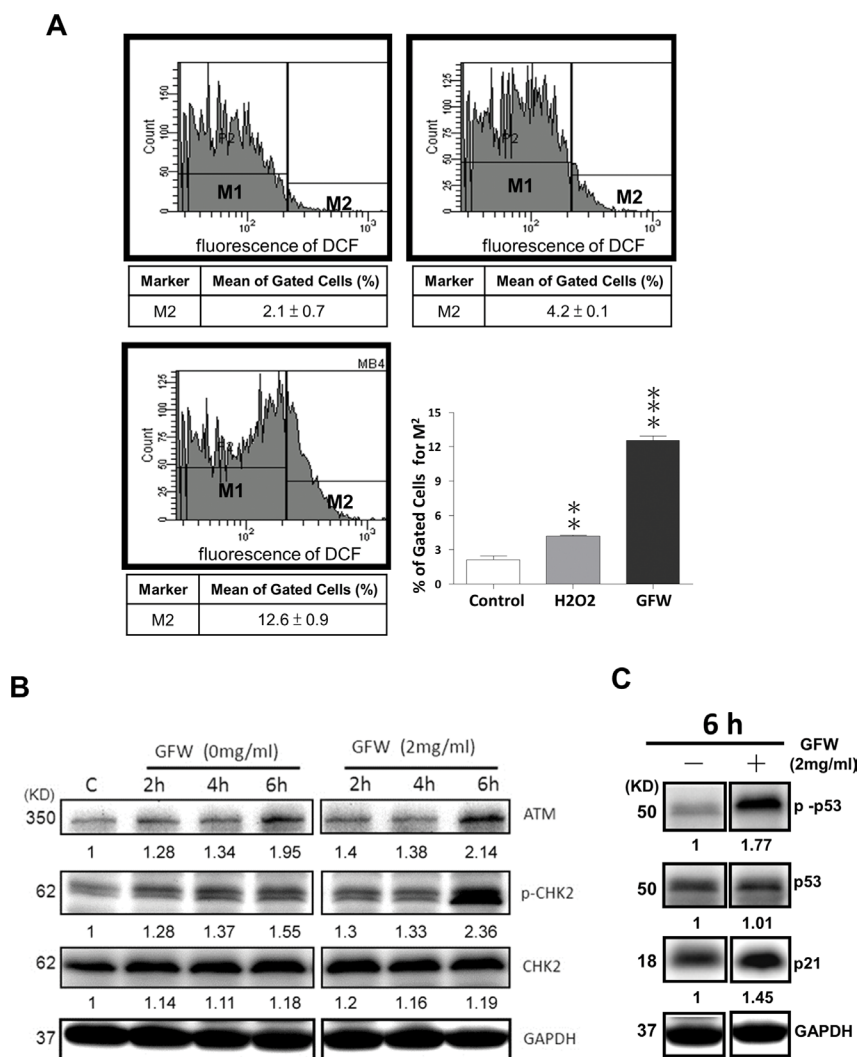


Figure 3. GFW enhanced ROS accumulation and activation of ATM/CHK2 and ATM/P53 signaling in MB49 cells. (A) MB49 cells were pre-incubated with DCFH-DA for 1 h, followed by treatment with 2 mg/mL GFW or 50 μ mol/L H_2O_2 for 3 h. Cells were then examined using flow cytometry to identify DCF fluorescent cells. Data are represented by M1 and M2 percentage of fluorescent cells. M1 is placed around the untreated cells. M2 is placed to the right of M1 as a designation of positive events. The mean \pm SEM ($n = 3$) of percentage gated for M2 is shown. Significant differences from the untreated controls are indicated by $**p \leq 0.01$ or $***p \leq 0.001$, as determined by one-way ANOVA and Dunnett comparison test. (B) MB49 cells untreated versus cells treated with GFW for 2, 4 and 6 h were examined for levels of ATM, CHK2 and Thr68-phosphorylated CHK2 by using Western blot analysis. (C) The expression levels of P21, P53 and Ser15-phosphorylated P53 were subsequently quantified in MB49 cells that did or did not undergo GFW treatment for 6 h. GAPDH protein was used as a loading control. Numbers indicate band intensities as determined by ImageJ software (National Institutes of Health; <http://rsb.info.nih.gov/ij/>).

in all mice treated with Mito-C or BCG throughout the study period, whereas only mild hematuria was observed in GFW-treated mice, which disappeared by d 17 (Table 1, Figure 4B). These results

indicate that GFW significantly reduced the gross/macroscopic hematuria caused by tumor growth. The monitoring of tumor growth was performed by using MRI on d 0, 4, 7 and 11 after GFW

treatment. On d 7, a number of tumors were observed in the animals that had not undergone any treatment (Figure 5, upper panel, C group), whereas fewer tumors were observed in the animal that did undergo treatment, even on the 11th day (Figure 5, lower panel, GFW group). Importantly, tumors treated with GFW presented ATM (Supplementary Figure S3) and p53 (Supplementary Figure S4) activation similar to that observed in cell line experiments. These findings demonstrate that GFW inhibited tumor growth *in vivo*.

On the final day of treatment, bladder volume was measured to compare the antitumor effects of intravesical treatment with GFW, BCG and mitomycin C. The mean bladder volumes in mice with or without intravesical PBS wash were 371.7 ± 335.1 and 445.1 ± 208 mm^3 , respectively. The mice treated with GFW, BCG and Mito-C presented mean bladder volumes of 95.1 ± 66 , 168.7 ± 123.3 and 51.9 ± 38.6 mm^3 , respectively (Figure 6A, Table 1). Note that the GFW, BCG and Mito-C treatment groups did not differ significantly with regard to mean bladder volume. All three groups presented volumes far lower than those in mice that did not undergo treatment, which is a clear indication that GFW is at least as effective as BCG and Mito-C in the inhibition of bladder tumor growth.

We subsequently used H&E staining for pathologic analysis of the urothelial wall of mouse bladders. As shown in Figure 6B, the urothelial wall of bladders in the GFW treatment group appear normal, whereas tumor cells can be observed in the control, BCG treatment, Mito-C treatment and PBS groups. Tumor cells were identified in approximately one in seven GFW-treated mice (14.3%), seven out of seven BCG-treated mice (100%) and two out of seven Mito-C-treated mice (28.6%) (Table 1), thereby demonstrating that intravesical treatment with GFW was the most effective treatment with regard to tumor suppression. Mouse kidney tissue was also subjected to pathologic analysis to determine whether metastasis or injury had occurred in the kidney after

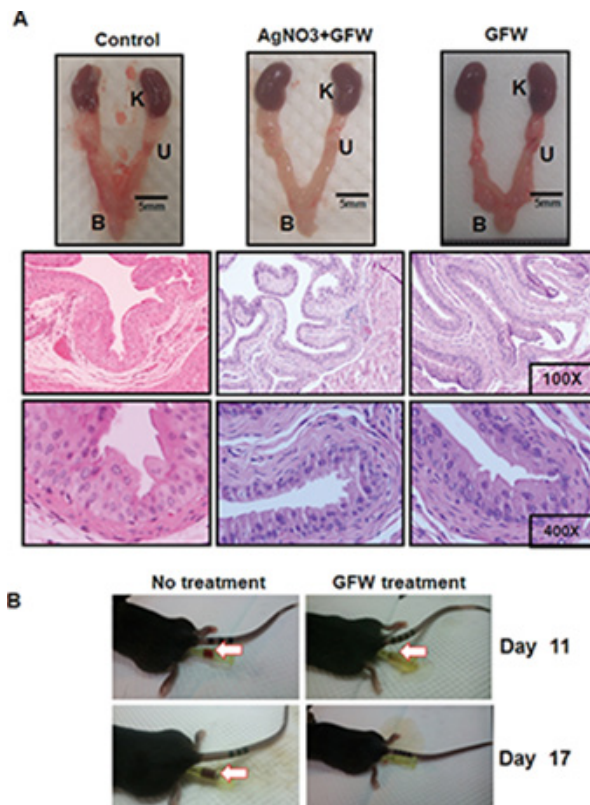


Figure 4. GFW is harmless to normal bladder urothelium of C57BL/6 mice and reduces the hematuria caused by tumor growth in mouse bladders. (A) Female C57BL/6 mice aged 6 wks with no tumor implantation underwent treatment with PBS or GFW (180 mg/mL) 10 times, administered every other day. "AgNO₃+GFW" indicates GFW-treated mice with initial AgNO₃-induced chemical lesions on bladder urothelium. After the mice were sacrificed, the urinary system was isolated and photographed (upper panel). K, U and B indicate the kidney, ureter and bladder, respectively. Bladder sections were stained with H&E for histopathology. Magnification: 100× (middle panel) and 400× (bottom panel). In all cases, the bladders presented a normal appearance and uniform urothelium, indicating that even 10 consecutive treatments with GFW was insufficient to harm the urothelium of normal mouse bladders. (B) After implantation of MB49 murine bladder cancer cells, urine was inspected and photographed. Serious events of hematuria were observed in all of the untreated control mice on d 11 and 19, while only mild hematuria was observed in GFW-treated mice on d 11. This result indicates that GFW reduced the gross hematuria caused by tumor growth in mouse bladder. The arrow indicates hematuria.

intravesical application of the drugs. Pathologic data failed to reveal tumor cells or lesions in any of the kidney sections (Supplementary Figure S5). These results indicate that metastasis had not occurred in our murine orthotopic bladder tumor model, indicating that intravesical treatment with these agents is harmless to the kidneys of orthotopic mice.

DISCUSSION

This study demonstrated the ability of GFW to suppress bladder cancer *in vitro* as well as *in vivo*. The anticancer activity of GFW appears to be associated with the inhibition of cancer cell growth via the induction of apoptosis and cell cycle arrest in the G1 phase. To reveal the effects of GFW on cell cycle progression in MB49 cells, we investigated the accumulation of intracellular ROS, the activation status of the ATM/CHK2 pathway and the expression levels of cell cycle regulatory protein P21. Our results indicated a correlation between GFW-induced G1 arrest in MB49 cells and P21 induction, which depends on ROS-induced activation of the ATM/CHK2 pathway (summarized in Figure 6C). This result is supported by the fact that intracellular ROS levels increased significantly in MB49 cells in response to GFW treatment for 3 h (Figure 3A). The increase in Thr68-phosphorylated CHK2 at 6 h, followed by an immediate increase in p21 expression, is a clear indication of the rapid, transient activation of ATM/CHK2 (Figure 3B). P21 is known to inhibit the activation of cyclin-dependent protein kinase

Table 1. Parameters in orthotopic bladder tumor model with and without intravesical therapy.

	Control	GFW	BCG	Mito-C	PBS	P ^a
Hematuria began (days postimplantation)	9.6 ± 1	9.3 ± 0.8	9.9 ± 0.4	9.0 ± 0	9.6 ± 1	0.1854
Hematuria ended (days postimplantation)	>22	17	>25	>22	>22	
Body weight (g) at the beginning	21.7 ± 2.5	21.9 ± 1.8	22.8 ± 3.4	20.7 ± 1.3	20.2 ± 1.6	0.2515
Body weight (g) at the end	19.0 ± 3.9	22.7 ± 1.2	20.5 ± 2.9	20.3 ± 1.6	19.7 ± 1.3	0.0774
Bladder volume (mm ³) at the end	445.1 ± 208	95.1 ± 66	168.7 ± 123.3	51.9 ± 38.6	371.7 ± 335.1	0.0012
Animals with tumor cells identified in bladder ^b	7 (7)	1 (7)	7 (7)	2 (7)	7 (7)	

^aANOVA.

^bNumber of test animals indicated in parentheses.

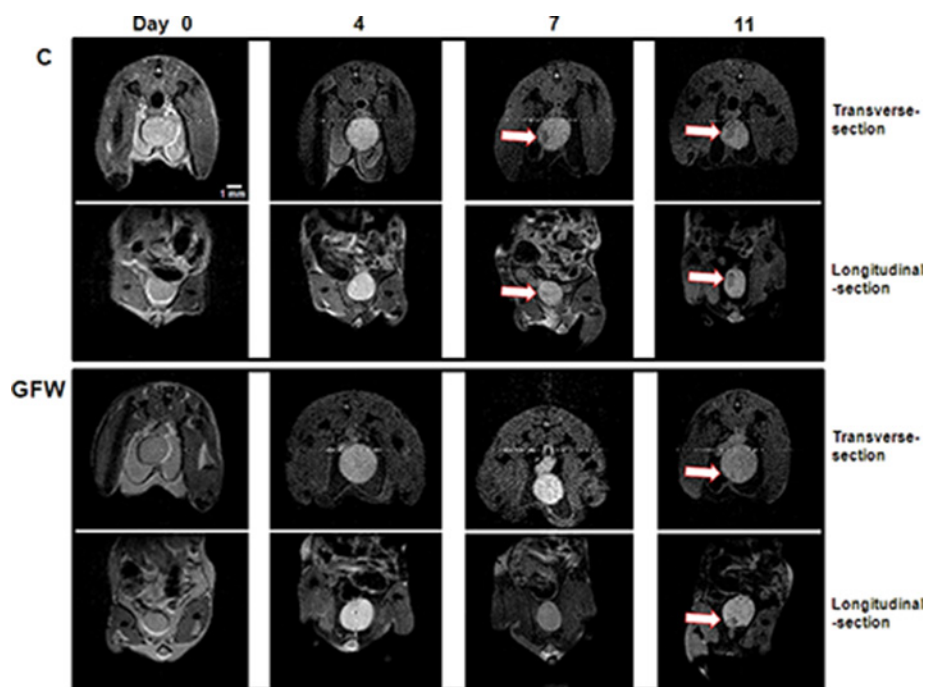


Figure 5. *In vivo* magnetic resonance detection of bladder tumor growth. Detection of bladder tumors was performed by using MRI on d 0, 4, 7 and 11 after implantation of murine bladder cancer cells. A transverse section and longitudinal section of the MRI results were photographed. These images show tumors in the untreated control mice observed on d 7 and 11 (upper panel, C group), while a relative small tumor was observed in the GFW-treated mice on d 11 (the lower panel, GFW group). This demonstrates the retardation of tumor growth by GFW. Arrows indicate bladder tumors. Scale bar, 1 mm.

Cdk2, leading to cell cycle arrest at the G1/S transition (35). Previous studies have also revealed the involvement of ATM/CHK2 signaling in the regulation of cell cycle progression in the G1/S phase (36) via the upregulation of P21 gene expression (37).

It has been demonstrated that cell cycle arrest can lead to the induction of apoptosis (38,39). In this study, MB49 cells consistently underwent GFW-induced apoptosis, as evidenced by the presence of a sub-G1 population (Figure 2A), a dose-dependent increase in apoptotic cells (Figure 2B) and an increase in the level of activated caspase-3 after treatment (Figure 2C). These findings are a clear indication that GFW is a potent inducer of apoptosis. ATM activation is responsible for the induction of ser15-phosphorylated P53 (40), leading to caspase-mediated

apoptosis (41). One current study has reported that Gypenosides, an active component isolated from the Chinese herb *Gynostemma pentaphyllum* (Thunb.) Makino, can induce apoptosis in human oral cancer SAS cells via ATM/ATR pathway activation, p53 phosphorylation and caspase-3 activation (42). Similarly, we detected an increase in the level of ATM, Ser15-phosphorylated p53 in MB49 cells after GFW treatment for 6 h (Figures 3B, C) and subsequent caspase-3 cleavage after GFW treatment for 32–48 h (Figure 2C). These results suggest that GFW induces the ATM/P53/caspase-3 signal transduction pathway, resulting in MB49 cell apoptosis (summarized in Figure 6C). Annexin V/propidium iodide flow cytometry analysis also revealed a significant increase in necrosis in MB49 cells after GFW treatment (Figure 2B). This result may be due to GFW-induced

activation of ATM/CHK2 signaling with the subsequent induction of P21 expression resulting in premature senescence. This scenario has previously been suggested to play a role in tumor suppression by reducing the replicative potential of cells (43). These effects are frequently detected in tumors obtained from patients who underwent genotoxic chemotherapy (44).

This study implemented a murine orthotopic bladder tumor model to analyze the *in vivo* effects of intravesical GFW therapy. In addition to inhibiting tumor growth in the bladder (Figure 5, Figure 6A), GFW also proved to be more effective than Mito-C and BCG as an intravesical agent in reducing hematuria associated with bladder tumor growth (Table 1). Notably, our pathologic data revealed that tumor cells appeared with far greater frequency in the BCG-treated group than in the Mito-C- and GFW-treated groups (100% versus 28.6% and 100% versus 14.3%, respectively; Figure 6B, Table 1), despite the similar effects of these three agents in inhibiting bladder tumor growth (Figure 6A). Interestingly, a formal meta-analysis of studies on comparative toxicity provided solid evidence to support the assertion that cystitis occurs more frequently among patients receiving BCG than among those administered Mito-C (53.8% versus 39.2%) (45). Nonetheless, we were unable to identify a significant number of lesions on mouse bladder tissue after 10 consecutive treatments with a high concentration of GFW (Figure 4A). This result is a demonstration of the low toxicity of GFW as an intravesical agent against bladder cancer. Moreover, intravesical therapy involves the local administration of the drug, thereby allowing direct contact between therapeutic agents and urothelial tumor. In cases where the bladder tumor is superficial, the direct application of GFW may provide more effective anticancer effects than would be possible with the oral administration of GFW. Finally, no cases of kidney

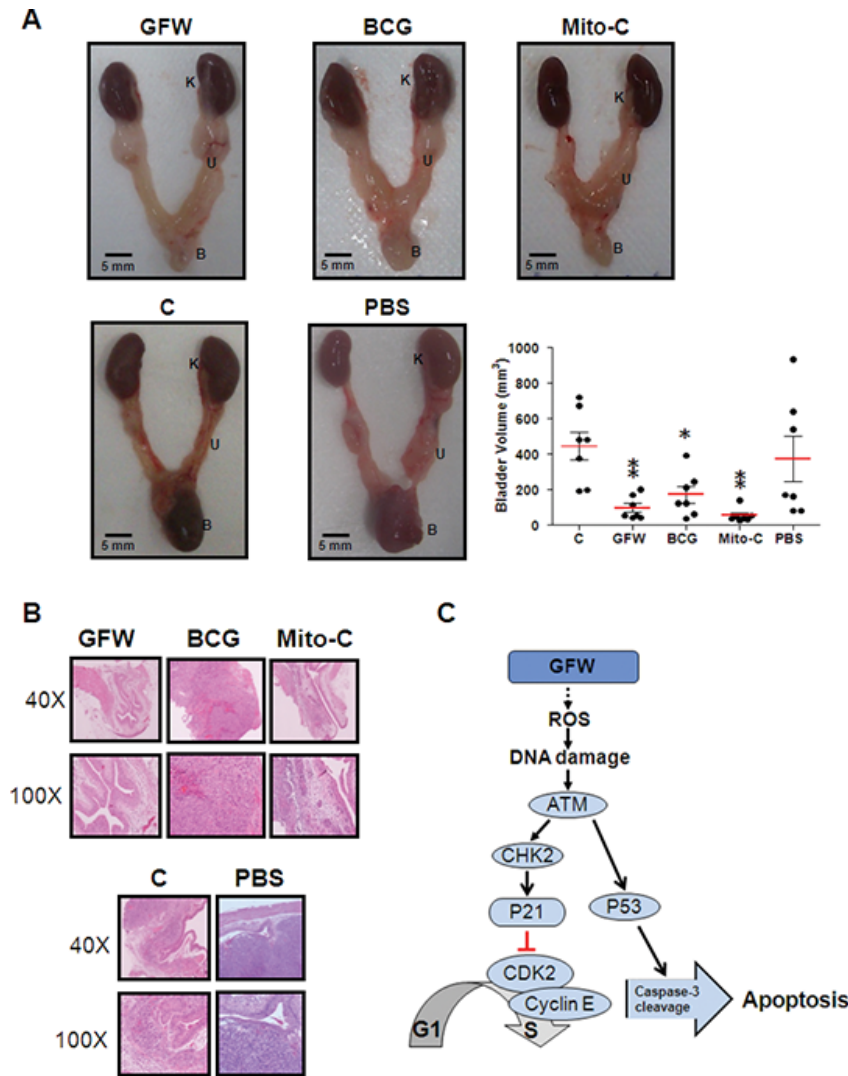


Figure 6. Effect of intravesical therapy in murine orthotopic bladder tumor model. (A) Female C57BL/6 mice aged 6 wks with tumor implantation were untreated (C) or treated with GFW, BCG, Mito-C or PBS according to the time schedule in Figure 1. After the mice were sacrificed, the urinary system was isolated and photographed. K, U and B indicate the kidney, urethra and bladder, respectively. The bladder volume was calculated by $(\text{length} \times \text{width}^2)/2$. Data are presented as mean \pm SEM ($n = 7$). Significant differences from the control are indicated by * $p < 0.05$, ** $p < 0.01$, as determined by one-way ANOVA and Dunnett comparison test. (B) After the treatment cycle, the mice were sacrificed and bladder sections were stained with H&E for histopathologic examination. Magnification: 40 \times (upper panel) and 100 \times (bottom panel). The urothelium in GFW-treated mice presented a normal uniform appearance. Tumor cells were observed only in the untreated control, BCG-treated, Mito-C-treated and PBS-treated mice. (C) Schematic diagram showing the effects of GFW on cell cycle arrest and apoptosis in MB49 murine bladder cancer cells. Intracellular ROS levels in MB49 cells presented a significant increase in response to GFW treatment with a subsequent rapid, transient activation of ATM/CHK2, followed by an immediate increase in P21 expression. P21 suppressed the activation of cyclin-dependent protein kinase Cdk2, leading to cell cycle arrest at the G1/S transition. In contrast, GFW induced apoptosis in murine bladder cancer MB49 cells via ATM activation, p53 phosphorylation and caspase-3 activation.

metastasis were observed in our murine orthotopic bladder tumor model, indicating that intravesical treatment with GFW, BCG or Mito-C is harmless to the kidneys of orthotopic mice (Supplementary Figure S2). Similar results were observed in the liver, lung and abdominal cavity of these mice.

CONCLUSION

The results of this study provide insight into the mechanisms underlying the effects of GFW on mouse bladder cancer cell MB49. We also evaluated the intravesicular application of GFW for the treatment of bladder cancer *in vivo*. Our data support the further investigation of GFW as a potential agent in the intravesical therapy of superficial bladder cancer.

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DISCLOSURE

The authors declare that they have no competing interests as defined by *Molecular Medicine*, or other interests that might be perceived to influence the results and discussion reported in this paper.

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